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Influence and reliability of lower-limb arterial occlusion pressure at different body positions.

Luke Hughes^{1,3}, Owen Jeffries¹, Mark Waldron¹, Benjamin Rosenblatt², Conor Gissane¹, Bruce Paton³, Stephen David Patterson¹

¹ School of Sport, Health and Applied Science, St Mary's University, London, UK, TW1 4SX*

² The Football Association. St. George's Park, Burton-Upon-Trent, UK

³ Institute of Sport, Exercise and Health, 170 Tottenham Court Road, London, UK

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Corresponding author: Dr Stephen Patterson

stephen.patterson@stmarys.ac.uk

* = full postal address

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1 **Abstract**

2

3 **Background:** Total arterial occlusive pressure (AOP) is used to prescribe pressures for
4 surgery, blood flow restriction (BFRE) and ischemic preconditioning (IPC). AOP is often
5 measured in a supine position; however, the influence of body position on AOP
6 measurement is unknown and may influence of the level of occlusion in different
7 positions during BFR and IPC. The aim of this study was therefore to investigate the
8 influence of body position on AOP. **Methods:** Fifty healthy individuals (age = 29 ± 6 y)
9 underwent AOP measurements on the dominant lower-limb in supine, seated and
10 standing positions in a randomised order. AOP was measured automatically using an
11 automated pneumatic tourniquet system, with each measurement separated by 5 minutes
12 of rest. **Results:** AOP was significantly lower in the supine position compared to the
13 seated position (187.00 ± 32.5 vs 204.00 ± 28.5 mmHg, $p < 0.001$) and standing position
14 (187.00 ± 32.5 vs. 241.50 ± 49.3 mmHg, $p < 0.001$). AOP was significantly higher in the
15 standing position compared to the seated position (241.50 ± 49.3 vs. 204.00 ± 28.5
16 mmHg, $p < 0.001$). **Discussion:** AOP measurement is body position dependent, thus for
17 accurate prescription of occlusion pressure during surgery, BFR and IPC, AOP should be
18 measured in the position intended for subsequent application of occlusion.

19

20 **Keywords:** Blood flow restriction exercise, Ischemic Preconditioning, Occlusion, Limb
21 occlusion pressure,

1 **Introduction**

2

3 The technique of occluding limb blood flow using pneumatic tourniquet cuffs is applied
4 in various settings, such as during surgery (Bussani & McEwen, 1988), blood flow
5 restriction exercise (BFRE) (Hughes et al., 2017) and ischemic preconditioning (IPC)
6 (Griffin et al., 2017). The level of occlusion achieved by an applied pressure is considered
7 to be an important factor for effective creation of a bloodless surgical field (Bussani &
8 McEwen, 1988), driving physiological adaptations and preventing full occlusion of
9 arterial blood flow during BFRE (Fahs et al., 2012; Lixandrão et al., 2015), and
10 effectiveness of the IPC stimulus (Cunniffe et al., 2016). The required pressure to reach
11 a desired level of occlusion is influenced by several factors, such as cuff width, limb
12 circumference and blood pressure (Loenneke et al., 2015; Jessee et al., 2016), which
13 makes standardisation of occlusion level difficult using arbitrary pressures. The use of
14 arbitrary pressures in BFRE may influence the amount of fatigue observed during
15 exercise and thus potential adaptations. Furthermore during IPC a standard pressure of
16 200 – 220 mmHg is widely used prior to exercise, irrespective of upper- or lower- limb
17 application, which may influence the potential benefit of this technique due to different
18 limb circumference and blood pressure within individuals (Bailey et al., 2012; Barbosa
19 et al., 2014; Patterson et al., 2015). Calculation of arterial occlusive pressure (AOP)
20 involves determination of the pressure required to fully occlude arterial flow to the
21 involved limb (AORN Recommended Practices Committee., 2007). This is most often
22 achieved using doppler ultrasound (Bezerra de Morais et al., 2017) and can be used to
23 prescribe pressure at a relative percentage of AOP to standardise the level of occlusion
24 across cohorts (Laurentino et al., 2012; Hughes et al., 2017; Patterson et al., 2017) .

25

26 Occlusion of blood flow is typically applied in one of three positions: supine, seated/semi-
27 recumbent or standing (Loenneke et al., 2012). However, within the literature it is evident
28 that only a small number of studies measure AOP in the same position that the occlusion
29 stimulus is subsequently applied (Staunton et al., 2015); this may not account for postural
30 influences on hydrostatic pressure (Wilkins, Halperin, & Litter, 1950; Eiken, 1988). For
31 example, movement of the lower-limb into a dependent position causes changes in
32 hydrostatic pressure, deformation of the vascular bed and an increase in blood flow and
33 pressure within the limb (Trinity et al., 2010). Systolic blood pressure has been identified
34 as a major predictive variable of AOP in the upper limbs (Loenneke et al., 2015); Jessee
35 et al., 2016), whereas thigh circumference is the largest predictor in the lower body
36 (Loenneke et al., 2015), thus it is conceivable that posture-induced changes in blood flow
37 and pressure may affect the pressure required for absolute occlusion of blood flow in that
38 limb.

39

40 To date, only one study has investigated the influence of body position on AOP
41 measurement (Sieljacks et al. 2018), however this was only in the supine and seated
42 position. Furthermore the reliability of AOP in each of these body positions is unknown.
43 This is problematic as it may result in under/over-estimation of the required pressure,
44 which may have implications for effectiveness in application of an occlusion stimulus.
45 Additionally, heterogeneous changes in AOP between individuals may lead to variation
46 in occlusion stimulus even when prescribed relative to AOP. Thus, the aim of this study
47 was to investigate the influence of different postural positions on AOP measurement. A
48 further aim was to examine the test-retest reliability of AOP at different body positions.

49

50 **Materials & Methods**

51

52 Participants

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54

55 Fifty participants (37 males and 13 females) volunteered to participate in the study.

56 Overall mean \pm SD for age, mass and stature were 29 ± 6 y, 77.3 ± 14.2 kg and $175.9 \pm$

57 8.4 cm, respectively. All were healthy, active non-smokers free from cardiovascular

58 (CV), pulmonary and metabolic diseases and musculoskeletal injuries in the past 12

59 months. Participants were asked to refrain from strenuous exercise, caffeine and alcohol

60 in the 24 hours prior to each testing session. All participants provided signed informed

61 consent in compliance with the Declaration of Helsinki, 7th version, October 2013 (World

62 Medical Association, 2013). All protocols were approved by St Mary's University ethical

63 committee (SMEC_2016-17_121).

64

65 Experimental design and procedure

66

67 To examine the influence of body position on AOP, participants attended the laboratory

68 on one occasion. Upon arrival, participant's mass and stature were recorded to the nearest

69 0.1 kg and 0.1 cm, respectively. Participants rested for 5 minutes in the supine position

70 on a portable treatment bed, then blood pressure was measured at the brachial artery

71 (Omron M5, Omron Healthcare, Europe B.V., the Netherlands, 14×48 cm). Thigh

72 circumference (cm) of the dominant lower-limb was measured at the midpoint of the

73 distance between the greater trochanter and the lateral condyle of the femur in accordance

74 with International Society for the Advancement of Kinanthropometry (ISAK) guidelines

75 using a flexible steel tape (Lufkin W606PM). Additionally, skinfold thickness (ST) was

76 measured (mm) at this point using skinfold calipers (Harpenden skinfold callipers, British

77 Indicators Ltd, UK). For the experimental procedure, participants underwent AOP
78 measurements in the dominant leg in a supine, seated and standing position in a within-
79 subjects randomised design. The randomisation was carried out by assigning each
80 participant a number and using publicly available software to allocate the order of
81 conditions (<http://www.randomization.com/>). Prior to each measurement, participants
82 rested in the required position for 5 minutes to ensure restoration of homeostasis after any
83 movement. For the supine position, participants lay on a portable treatment bed with their
84 arms relaxed by their sides. For the seated position, participants sat upright with their legs
85 straight and the hip flexed at a 90° angle, assessed using a goniometer. For the standing
86 position, participants stood in the standard anatomical position. Prior to each
87 measurement, participants rested in the required position for 5 minutes to ensure
88 restoration of normal blood flow after any movement (Jessee et al., 2016).

89 AOP measurement

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91
92 Restriction of blood flow in the lower-limb was achieved using the Delfi Easy-fit variable
93 contour tourniquet cuff (11.5 cm x 86 cm x 5 mm), connected to a pneumatic cuff inflator
94 (Delfi PTS, Delfi Medical, Vancouver, BC, Canada). The pneumatic tourniquet was
95 equipped with the capability of automatically measuring AOP and calculating the
96 personalised tourniquet pressure, comprised of a dual-purpose personalised tourniquet
97 cuff and a personalised tourniquet instrument containing AOP calculation sensors and
98 software. The pneumatic system connected to the tourniquet cuff increased the cuff
99 pressure in stepwise increments, analysing the pneumatic pressure pulsations in the cuff
100 bladder by the arterial pressure pulsations at each cuff pressure increment, and used these
101 characteristics to determine AOP (McEwen et al., 2015). AOP measurement using this

102 cuff was found to not be clinically or statistically different from using the gold standard
103 doppler technique (+0.08 mmHg [95%CI -2.66 to 2.82] for lower limbs) across 257 pairs
104 of AOP measurements taken from upper and lower limbs in 143 pre- and post-surgical
105 patients aged 17-86 (McEwen et al., 2015; Masri et al., 2016). This technique of
106 measuring AOP was found to have clinically acceptable accuracy compared to the distal-
107 sensor-based method of automatic AOP measurement, which measures AOP using a
108 sensor located on the most distal phalange of the involved limb (McEwen, Inkpen, &
109 Younger, 2002). The variable-contour cuff was placed on the most proximal portion of
110 the participant's dominant lower limb directly onto the skin and connected to the
111 pneumatic tourniquet with airtight hose tubing. After 5 minutes of rest, the device was
112 turned on to calculate AOP in the manner described above. The AOP displayed on the
113 pneumatic tourniquet device was recorded for each of the three positions.

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Test-retest reliability

118 To assess the reliability of the pneumatic tourniquet, 10 subjects visited the laboratory at
119 the same time of day, on a second occasion, one week later during which the experimental
120 procedure for the AOP measurement was repeated with the order of positions tested in
121 the same order as they had previously been tested.

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Statistical analysis

126 Due to non-normal distribution of the supine body position data ($p < 0.05$) which persisted
127 after log transformations, a non-parametric Friedman test was used to determine if there
128 were differences in AOP across the three different body positions. For any significant
129 differences, Wilcoxon signed-rank pairwise comparisons were performed with

130 Bonferroni correction. Within-subject coefficient of variation (COV) was calculated, and
131 an intraclass correlation coefficient (ICC) test with a two-way mixed effects model was
132 used to determine absolute agreement to examine the reliability and reproducibility of
133 AOP measurements with the pneumatic tourniquet system. The COV was calculated from
134 the ratio of the standard deviation (SD) and the mean of the two AOP measurements (CV
135 $= ((SD/mean) \times 100)$), followed by calculation of the mean (Bezerra de Moraes et al.,
136 2017).

137

138 **Results**

139

140 **Participants**

141

142 All 50 participants completed the study with no adverse events. Participants' blood
143 pressure, resting heart rate, thigh circumference and ST are presented in Table 1.

144

145 ***Insert Table 1 here***

146

147 **AOP**

148

149 Data are presented as mean \pm SD. AOP was statistically significantly different in the
150 different body positions, $\chi^2(2) = 90.04$, $p < 0.001$. Post-hoc analysis revealed that AOP in
151 the supine position was significantly lower compared to the seated position ($187.00 \pm$
152 32.5 vs 204.00 ± 28.5 mmHg, respectively, $p < 0.001$), and the standing position (187.00
153 ± 32.5 vs 241.50 ± 49.3 mmHg, respectively, $p < 0.001$). Additionally, AOP in the seated
154 position was significantly lower than in the standing position (204.00 ± 28.5 vs $241.50 \pm$
155 49.3 mmHg, respectively, $p < 0.001$).

156

157 Test-retest reliability

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159

160 For the supine position, the ICC for assessing reliability of the device across two repeated

161 measurements was 0.982 (95% CI: 0.932 to 0.995) with a COV of 2.94% (95% CI: 1.90

162 to 3.98%). For the seated position, the ICC for assessing reliability of the device across

163 two repeated measurements was 0.975 (95% CI: 0.932 to 0.994) with a COV of 1.82%

164 (95% CI: 0.95 to 2.69). For the standing position, the ICC for assessing reliability of the

165 device across two repeated measurements was 0.953 (95% CI: 0.822 to 0.988) with a

166 COV of 2.97% (95% CI: 0.89 to 5.05).

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171

172 **Discussion**

173

174 This study investigated the influence of body position on AOP measurement. The main

175 findings were that lower-limb arterial AOP is body-position dependent. For absolute

176 occlusion of lower -limb arterial blood flow, it appears that higher pressures are required

177 in a seated compared to supine body position, and higher pressures are required in a

178 standing compared to seated and supine body position.

179

180 The pressure required to fully restrict arterial blood flow to the lower-limb increased from

181 187 mmHg to 204 mmHg to 241 mmHg in the supine, seated and standing positions,

182 respectively. This reflects literature demonstrating increases in peripheral blood flow to

183 the extremities (Goetz, 1950) and changes in hydrostatic pressure with different body
184 positions (Wilkins et al., 1950; Eiken, 1988). Elevation of the heart above the limbs when
185 comparing an seated to supine position results in an increase in peripheral blood flow and
186 pooling of blood in the lower limbs due to gravitational forces (Olufsen et al., 2005).
187 Increases in lower-limb local hydrostatic pressure (Wilkins et al., 1950), mechanical
188 deformation of the vascular bed and stimulation of group III afferent fibres (Trinity et al.,
189 2011) triggers peripheral vasodilation, causing a rise in peripheral blood flow. As there
190 is greater elevation of the heart again in a standing position and a larger effect of gravity,
191 these changes in peripheral blood flow and pressure may be amplified further in a
192 standing position (Olufsen et al., 2005). Studies examining factors influencing AOP in
193 the upper-limb support this, with systolic blood pressure identified as one of the major
194 predictive variables of AOP in the upper-limb (Loenneke et al., 2014; Jessee et al., 2016).

195

196 Implications for BFRE

197

198 These observations suggest that measurement of AOP and application of the occlusive
199 stimulus in different positions would result in undesirable levels of occlusion, which has
200 important implications for application. For example, if AOP is measured whilst standing
201 but occlusion is applied in a seated or supine position, the individual may be exposed to
202 higher levels of occlusion than necessary. Within BFRE, higher pressures have been
203 shown to cause greater CV responses to exercise (Rossow et al., 2012) and may result in
204 full restriction of arterial inflow to the working muscle. It has been speculated that this
205 may increase the risk of ischemic reperfusion injury, peripheral nerve injury or
206 concerning hemodynamic alterations (Kacin et al., 2015, Loenneke et al., 2011, Jessee et

207 al., 2016), particularly when BFRE is used in patients with blood-related conditions such
208 as hypertension and heart disease (Madarame et al., 2010; Cezar et al., 2016).
209 Additionally, although the focus of this study was not on the physiological responses to
210 BFR, higher pressures are known to increase discomfort responses to BFRE (Jessee et
211 al., 2017; Mattocks et al., 2017) and thus could impact upon the clinical utility of BFRE
212 training and patient adherence to clinical rehabilitation programmes Therefore, accurate
213 calculation of AOP for pressure prescription is required for selection of the minimum
214 occlusion pressure required to elicit a positive change. It is of note that optimal occlusion
215 pressure is not fully understood, and may vary in different contexts. However, current
216 literature suggests that light-load BFRE (<30% 1RM) training protocols benefit from
217 higher occlusion pressures (80% vs 40%) (Lixandrão et al., 2015), which would support
218 the importance of accurate AOP measurement for prescription of relative pressures. In
219 contrast when loads are $\geq 30\%$ 1RM there does not appear to be a need to exercise at
220 higher percentages of AOP (Counts et al., 2016).

221

222 On the contrary, measurement of AOP in a supine position and subsequent application of
223 BFRE in a seated or standing position may result in a lower level of occlusion than
224 desired, or a lack of venous occlusion altogether in situations where low pressures are
225 used (Kubota et al., 2011). Furthermore higher pressures during BFRE result in greater
226 accumulation of metabolic byproducts (Yasuda et al., 2010) hypothesised to be one of the
227 major driving forces of hypertrophic adaptations to light load BFR training (Pearson &
228 Hussain, 2015; Hughes et al., 2017). Insufficient levels of restriction due to inaccurate
229 pressure prescription confounded by body position may reduce the metabolic stress
230 stimulus, which may dampen the hypertrophic BFR stimulus and partially explain reports

231 of ineffectiveness of BFRE. Furthermore AOP may be influenced by time of day, with
232 increased pressure observed as the day progresses, likely brought about by oscillatory
233 changes in changes in blood flow and pressure ((Ingram et al., 2017). Therefore
234 measurement position and time of day should be considered by those using BFRE over
235 repeated applications.

236

237 Implications for IPC

238

239 When performing IPC before exercise, the lower or upper limb is occluded at a set
240 arbitrary pressure between 200 and 220mmHg (Bailey et al., 2012; Barbosa et al., 2014;
241 Patterson et al., 2015). Furthermore when applying this pressure, participants are either
242 supine (Patterson et al 2015) or in a seated position (Marocolo et al., 2017) which may
243 influence the amount of occlusion observed. In the current study the average pressure
244 observed in the supine position was < 200 mmHg, however in the seated position this
245 increased to 204 mmHg. This suggests that the normal arbitrary pressures of 220 mmHg
246 should be sufficient to fully restrict blood flow prior to this intervention. However in
247 some studies the pressure used has been 200mmHg (Barboasa et al., 2014) and as low as
248 180 mmHg (Cunniffe et al., 2016). In the current investigation 28% and 18% of the
249 participants would not be fully occluded at 200 and 220mmHg in the supine position,
250 respectively. Furthermore, in the seated position, this number would rise to 60% and 28%
251 for 200 and 220 mmHg, respectively. Therefore we recommend the use of AOP to
252 standardise pressures during IPC due to the wide variance in participants and also the
253 wide array of cuffs used to occlude individuals.

254

255 Reliability

256 When measuring AOP automatically, it is important that the device used is reliable and
257 consistent across repeated measures to ensure correct prescription of pressure. In this
258 study, the pneumatic tourniquet system appeared to have high reproducibility (> 0.953)
259 with a COV of less than 2.97% across all the body positions examined. These findings
260 are similar to a recent study examining the reliability of doppler ultrasound for calculating
261 total AOP in the upper limbs (Bezerra de Morais et al., 2017). The authors calculated
262 AOP using doppler ultrasound in 13 male volunteers across three repeated measures,
263 reporting an ICC of 0.795 and a COV of 5.6%. Although the present study was in the
264 lower-limbs, we observed greater ICC scores and smaller COVs, suggesting
265 measurement of AOP using the pneumatic tourniquet system may be more reliable than
266 doppler ultrasound. This is may be attributed to the absence of human error; however,
267 this is speculative at present. Nevertheless, other studies have demonstrated similar
268 results.. The results of the present study suggest the pneumatic tourniquet system is highly
269 reproducible for measuring lower-limb arterial AOP due to the high ICC values and lower
270 COV scores compared to similar studies in the upper- limb (Bezerra de Morais et al.,
271 2017).

272

273

274

275 **Conclusion**

276 The findings of the present study have several important clinical implications. Firstly, it
277 appears that AOP is body position-dependent. In BFRE and IPC, AOP must therefore be
278 calculated in the position of exercise to ensure accurate occlusion, while minimising the

279 risk of an adverse CV/neurological event or application of an insufficient BFRE stimulus.
280 Secondly, it appears that the pneumatic tourniquet system can be used to reliably calculate
281 lower-limb AOP. Previously, we highlighted that AOP may change across the duration
282 of a BFRE training study due to various tissue adaptations, such as increases in muscle
283 mass and vasculature adaptations, thus it is important to continually monitor AOP to
284 ensure prescription of the correct pressure (Hughes et al., 2017). Doppler ultrasound tools
285 can be expensive, and repeated measurement of AOP using this technique prior to weekly
286 BFRE training sessions would likely be time-consuming and require considerable skill.
287 This may be exacerbated in a clinical rehabilitation setting such as the NHS where
288 rehabilitation exercise classes are already time-constrained. However, the pneumatic
289 tourniquetsystem provides a simple and quick alternative for calculating AOP, and may
290 be implemented on a session-to-session basis. We propose that an actual measurement of
291 AOP is obtained at rest, prior to BFRE, and a percentage of that measurement is used
292 provide a more reliable stimulus (Laurentino et al., 2012; Hughes et al., 2017; Patterson
293 et al., 2017) as this method is still under-utilised by practitioners (Patterson & Brandner
294 2017).

295

296 In conclusion, the results of this study indicate that for accurate prescription of occlusion
297 pressure in BFRE and IPC applications, body position must be accounted for an AOP
298 measured in the position that the occlusion stimulus will be subsequently applied.
299 Moreover, the pneumatic tourniquet system appears to have high reproducibility for
300 automatic measurement of AOP in the lower-limbs.

301

302 **Acknowledgments**

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304 for this study.

305

306 **Conflicts of interest**

307 None.

References

AORN Recommended Practices Committee. Recommended practices for the use of the pneumatic tourniquet in the perioperative practice setting. 2007 AORN Journal 86:640–655.

Bailey TG, Jones H, Gregson W, Atkinson G, Cable NT, Thijssen DH. Effect of ischemic preconditioning on lactate accumulation and running performance. 2012. *Medicine & Science in Sports & Exercise* 44:2084–9.

Barbosa T, Machado AC, Braz ID, Fernandes IA, Vianna LC, Nobrega AC, Silva BM. Remote ischemic preconditioning delays fatigue development during handgrip exercise. 2015. *Scandinavian Journal of Medicine & Science in Sports* 25:356–642014;

Bezerra de Moraes AT, Santos Cerqueira M, Moreira Sales R, Rocha T, Galvão de Moura Filho A. Upper limbs total occlusion pressure assessment: Doppler ultrasound reproducibility and determination of predictive variables. 2017. *Clinical Journal of Physiology & Functional Imaging* 37:437–441.

Bussani CR, McEwen JA. Improved tracking of limb occlusion pressure for surgical tourniquets. 1988. *IEEE Translational & Biomedical Engineering* 35: 221-229.

Cezar MA, De Sá CA, Corralo V da S, Copatti SL, Santos GAG dos, Grigoletto ME da S. 2016. Effects of exercise training with blood flow restriction on blood pressure in medicated hypertensive patients. *Motriz: Revista de Educação Física* 22:9–17.

Counts BR, Dankel SJ, Barnett BE, Kim D, Mouser JG, Allen KM, Thiebaud RS, Abe

T, Bemben MG, Loenneke JP. 2016. Influence of relative blood flow restriction pressure on muscle activation and muscle adaptation. *Muscle & nerve*, 53:438-445.

Cunniffe B, Sharma V, Cardinale M, Yellon D. Characterization of muscle oxygen response to vasvular occlusion: implications for remote ischaemic preconditioning and physical performance. 2017. *Clinical Journal of Physiology & Functional Imaging* 37; 785-793.

Eiken O. Effects of increased muscle perfusion pressure on responses to dynamic leg exercise in man. 1988. *European Journal of Applied Physiology Occupational Physiology* 57:772-6.

Fahs CA, Loenneke JP, Rossow LM, Thiebaud RS, Bemben MG. Methodological considerations for blood flow restricted resistance exercise. 2012. *Journal of Trainology* 1:14-22.

Goetz RH. Effect of Changes in Posture on Peripheral Circulation, with Special Reference to Skin Temperature Readings and the Plethysmogram. 1950. *Circulation* 1:56-75.

Griffin PJ, Ferguson RA, Gissane C, Bailey SJ, Patterson SD. Ischemic preconditioning enhances critical power during a 3 minute all-out cycling test. 2017. *Journal of Sports Science* doi.org/10.1080/02640414.2017.1349923.

Hughes L, Paton B, Rosenblatt B, Gissane C, Patterson SDS. Blood flow restriction training in clinical musculoskeletal rehabilitation: a systematic review and meta-

- analysis. 2017. *British Journal of Sports Medicine* 51:1003–1011.
- Ingram JW, Dankel SJ, Buckner SL, Counts BR, Mouser JG, Abe T, Laurentino GC, Loenneke JP. 2017. The influence of time on determining blood flow restriction pressure. *Journal of science and medicine in sport*, 20:777-780.
- Jessee MB, Dankel SJ, Buckner SL, Mouser JG, Mattocks KT, Loenneke JP. The Cardiovascular and Perceptual Response to Very Low Load Blood Flow Restricted Exercise. 2017. *International Journal of Sports Medicine* 38:597–603.
- Jessee MB, Buckner SL, Dankel SJ, Counts BR, Abe T, Loenneke JP. The Influence of Cuff Width, Sex, and Race on Arterial Occlusion: Implications for Blood Flow Restriction Research. 2016. *Sport Medicine* 46:913–921.
- Kacin A, Žargi TG, Rosenblatt B, Biswas A. Safety Considerations With Blood Flow Restricted Resistance Training. 2015. *Annales Kinesiologiae* 6:3–26.
- Kubota A, Sakuraba K, Koh S, Ogura Y, Tamura Y. Blood flow restriction by low compressive force prevents disuse muscular weakness. 2011. *Journal of Science and Medicine in Sport* 14:95–99.
- Laurentino G, Ugrinowitsch C, Roschel H, Aoki MS, Soares AG, Neves M, Aihara AY, da Rocha Correa Fernandes A, Tricoli V. Strength training with blood flow restriction diminishes myostatin gene expression. 2012. *Medicine and Science in Sports and Exercise* 44. 406-412.
- Lixandrão ME, Ugrinowitsch C, Laurentino G, Libardi CA, Aihara AY, Cardoso FN, Tricoli V, Roschel H. Effects of exercise intensity and occlusion pressure after

- 12 weeks of resistance training with blood-flow restriction. 2015. *European Journal of Applied Physiology* 115:2471–2480.
- Loenneke JP, Allen KM, Mouser JG, Thiebaud RS, Kim D, Abe T, Bembem MG (2015) Blood flow restriction in the upper and lower limbs is predicted by limb circumference and systolic blood pressure. 2015. *European Journal of Applied Physiology* 2:397–405.
- Loenneke JP, Allen KM, Mouser JG, Thiebaud RS, Kim D, Abe T, Bembem MG. Blood flow restriction in the upper and lower limbs is predicted by limb circumference and systolic blood pressure. 2014. *European Journal of Applied Physiology* 115:397–405.
- Loenneke JP, Fahs CA, Rossow LM, Sherk VD, Thiebaud RS, Abe T, Bembem DA, Bembem MG. Effects of cuff width on arterial occlusion: Implications for blood flow restricted exercise. 2012. *European Journal of Applied Physiology* 112:2903–2912.
- Loenneke JP, Wilson JM, Wilson GJ, Pujol TJ, Bembem MG. Potential safety issues with blood flow restriction training. 2011. *Scandinavian Journal of Medicine and Science in Sport* 21:510–518.
- Madarame H, Kurano M, Takano H, Iida H, Sato Y, Ohshima H, Abe T, Ishii N, Morita T, Nakajima T. Effects of low-intensity resistance exercise with blood flow restriction on coagulation system in healthy subjects. 2010 *Clinical Physiology and Functional Imaging* 30:210–213.
- Marocolo IC, da Mota GR, Londe AM, Patterson SD, Barbosa Neto O, Marocolo M.

Acute ischemic preconditioning does not influence high-intensity intermittent exercise performance 2017 PEER J. Nov 30;5:e4118. doi: 10.7717/peerj.4118

Mattocks KT, Jessee MB, Counts BR, Buckner SL, Grant Mouser J, Dankel SJ, Laurentino GC, Loenneke JP. The effects of upper body exercise across different levels of blood flow restriction on arterial occlusion pressure and perceptual responses. 2017. *Physiology & Behavior* 171:181–186.

Masri BA, Day B, Younger ASE, Jeyasurya J. Technique for Measuring Limb Occlusion Pressure that Facilitates Personalized Tourniquet Systems: A Randomized Trial. *J Med Biol Eng* 2016;36:644–650

McEwen JA, Masri BA, Day B, Younger AS. Development of Personalised Tourniquet Systems using a new technique for measuring limb occlusion pressure. 2015. *IFMBE Proceedings* 51.

McEwen JA, Inkpen KB, Younger A. Thigh tourniquet safety: Limb occlusion pressure measurement and a wide contoured cuff allow lower cuff pressure. 2002. *Surgical Technology* 34:8–18

Olufsen MS, Ottesen JT, Tran HT, Ellwein LM, Lipsitz LA, Novak V. Blood pressure and blood flow variation during postural change from sitting to standing: model development and validation. 2005. *Journal of Applied Physiology* 99:1523–37.

Patterson SD, Brandner C. The role of blood flow restriction training for applied practitioners: A questionnaire based survey. 2017. *Journal of Sports Science* 36: 123-130.

- Patterson SD, Hughes L, Head P, Warmington S, Brandner C, Blood flow restriction training: A novel approach to augment clinical rehabilitation: how to do it. 2017. *British Journal of Sports Medicine* 51:1648-1649.
- Patterson SD, Bezodis NE, Glaister M, Pattison JR. The effect of ischemic preconditioning on repeated sprint cycling performance. 2015. *Medicine and Science in Sports and Exercise* 47:1652–8
- Pearson SJ, Hussain SR. A Review on the Mechanisms of Blood-Flow Restriction Resistance Training-Induced Muscle Hypertrophy. 2015. *Sport Medicine* 45:187–200.
- Rossow LM, Fahs CA, Loenneke JP, Thiebaud RS, Sherk VD, Abe T, Bemben MG. Cardiovascular and perceptual responses to blood-flow-restricted resistance exercise with differing restrictive cuffs. 2012. *Clinical Journal of Physiology and Functional Imaging* 32:331–337.
- Sieljacks S, Knudsen L, Wernbom M, Vissing K. Body position influence arterial occlusion pressure: implications for the standardization of pressure during blood flow restricted exercise. 2018. *European Journal of Applied physiology* doi.org/10.1007/s00421-017-3770-2.
- Staunton CA, May AK, Brandner CR, Warmington SA. Haemodynamics of aerobic and resistance blood flow restriction exercise in young and older adults. 2015. *European Journal of Applied Physiology* 115:2293–2302.
- Trinity JD, McDaniel J, Venturelli M, Fjeldstad AS, Ives SJ, Witman MAH, Barrett-O’Keefe Z, Amann M, Wray DW, Richardson RS. Impact of body position on

central and peripheral hemodynamic contributions to movement-induced hyperemia: implications for rehabilitative medicine. 2011. *AJP Heart & Circulatory Physiology* 300:H1885–H1891.

Trinity JD, Amann M, McDaniel J, Fjeldstad AS, Barrett-O’Keefe Z, Runnels S, Morgan DE, Wray DW, Richardson RS. Limb movement-induced hyperemia has a central hemodynamic component: evidence from a neural blockade study. 2010. *AJP Heart & Circulatory Physiology* 299:H1693–H1700.

Wilkins RW, Halperin MH, Litter J. The Effect of the Dependent Position upon Blood Flow in the Limbs. 1950. *Circulation* 11:373–379.

World Medical Association. World Medical Association Declaration of Helsinki. Ethical principles for medical research involving human subjects. 2013. *JAMA* 310:2191–2194.

Yasuda T, Abe T, Brechue WF, Iida H, Takano H, Meguro K, Kurano M, Fujita S, Nakajima T. 2010. Venous blood gas and metabolite response to low-intensity muscle contractions with external limb compression. *Metabolism: clinical and experimental*, 59:1510-1519.

Table legends

Table 1 Participant anthropometric characteristics (Mean \pm SD)