

**Title: Hypertonic Saline-Evoked Muscle Pain in the Quadriceps Reduces Neuromuscular Performance and Alters Corticospinal Excitability**

**Running Head:** Muscle pain and neuromuscular function

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## 19   **Abstract**

20   Muscle pain can alter corticospinal function, but the specific excitatory/inhibitory effects on the  
21   quadriceps across different levels of corticospinal neuron recruitment remain unclear. Furthermore,  
22   maximal force production is reduced with muscle pain, but how the rate of force development, a key  
23   component of neuromuscular function remains less-known. To investigate this, healthy participants  
24   completed an isometric maximal voluntary contraction (MVC) followed by submaximal, intermittent  
25   contractions after receiving a hypertonic saline injection into the vastus lateralis to cause quadriceps  
26   pain (HYP) or isotonic saline, a non-painful control (ISO). Peripheral nerve stimulation was delivered  
27   during and after MVCs to determine neuromuscular function. Transcranial magnetic stimulation  
28   (TMS) was delivered at 120% and 150% of active motor threshold during submaximal contractions to  
29   determine corticospinal excitability/inhibition, along with paired-pulse TMS to determine short-  
30   interval intracortical inhibition (SICI). Results revealed a moderate effect size (ES) reduction in MVC  
31   force ( $ES = -0.68$ ,  $P = 0.020$ ), early-phase rate of force development ( $ES = -0.57$ ,  $P = 0.029$ ), and  
32   voluntary activation ( $ES = -0.66$ ,  $P = 0.008$ ) in HYP compared to ISO. Corticospinal excitability  
33   increased in HYP compared to ISO ( $ES = 0.60$ ,  $P = 0.023$ ), whereas corticospinal inhibition decreased  
34   in HYP at higher stimulation intensities only ( $ES = 0.63$ ,  $P = 0.017$ ). Conversely, SICI increased in  
35   HYP compared to ISO ( $ES = 0.58$ ,  $P = 0.035$ ). Our findings indicate that muscle pain induced by a  
36   hypertonic saline injection reduced quadriceps neuromuscular function due to centrally mediated  
37   mechanisms, potentially involving both excitatory and inhibitory effects on the corticospinal tract.

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45    **New and Noteworthy:**

46    Hypertonic saline-induced quadriceps muscle pain reduced knee-extensor maximal voluntary  
47    force, rate of force development and voluntary activation, without altering peripheral muscle  
48    function, suggesting a centrally mediated impairment of neuromuscular performance in  
49    healthy individuals. Alongside these changes was an increase in corticospinal excitability at  
50    both low and high stimulation intensities, whereas pain decreased corticospinal inhibition at  
51    high stimulation intensities only. Furthermore, hypertonic saline-induced pain increased  
52    intracortical inhibition, suggesting non-uniform effects of pain on the corticospinal tract.

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## 68 **Introduction**

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70       Muscle pain is a salient and widespread experience in health and disease. Pain can be  
71 defined as ‘an unpleasant sensory and emotional experience associated with, or resembling  
72 that associated with, actual or potential tissue damage’ (1). Clinically, individuals with  
73 conditions such as fibromyalgia and complex regional pain syndrome can display elevated  
74 levels of appendicular muscle pain (1–3). These conditions are also associated with impaired  
75 neuromuscular function and reduced voluntary activation, as highlighted in recent meta-  
76 analyses (4, 5). The manifestation of muscle pain is also a common experience both during  
77 exercise (exercise-induced pain) (6) or in the days following intense or unaccustomed  
78 exercise in the form of mechanical hyperalgesia, commonly referred to as delayed onset  
79 muscle soreness (7). These pain scenarios typically arise from stimulation of group III/IV  
80 nociceptive afferents by noxious concentrations of biochemicals associated with anaerobic  
81 energy contribution and inflammation (e.g., hydrogen ions, adenosine, potassium) (8–10).  
82 Nociceptive signals transmitted from group III/IV afferents synapse on to dorsal horn of the  
83 spinal cord, where they ascend to several brain areas including the primary somatosensory  
84 cortex, resulting in the perception of pain (6).

85       In addition to generating a conscious pain perception, nociceptive signals can also  
86 impact the function of the primary motor cortex and the corticospinal pathway (11). This is of  
87 particular interest because motor performance of the lower limb (primarily the quadriceps  
88 femoris), relies on the corticospinal pathway, as it controls voluntary movement, coordinates  
89 muscle activation, integrates sensory feedback, and adapts to pain or fatigue (11). Altered  
90 function may have negative implications for both exercise performance success and  
91 completing activities of daily living (e.g., stair climbing).

92       A consistently observed consequence of acute muscle pain in the lower limb is a  
93 reduction in maximal voluntary force production (12–15). Submaximal forces up to 80% of

94 maximum can be produced in the lower limbs during acute muscle pain, albeit with reduced  
95 endurance capacities (14, 16–19) and a greater perceived effort (6, 14). However, the impact  
96 of acute muscle pain on the ability to produce submaximal forces rapidly (i.e., rate of force  
97 development [RFD]) is unclear. Given that RFD is functionally relevant for tasks requiring  
98 rapid force generation, such as sprinting, jumping, or reacting to sudden changes in the  
99 environment (20), there is a need to determine how acute muscle pain influences this critical  
100 aspect of motor performance.

101 Mechanisms underpinning these motor performance changes during acute muscle  
102 pain have also been investigated using various neurophysiological measurement techniques.  
103 The interpolated twitch technique has been utilised to identify changes in voluntary activation  
104 during maximal voluntary contractions (21), with consistent decreases in voluntary activation  
105 observed during experimental quadriceps pain, suggesting that central mechanisms are  
106 responsible for decrements in physical task performance (14, 22, 23). However, current  
107 theory proposes that pain can have both excitatory and inhibitory effects on the  
108 neuromuscular system which serves as a protective mechanism to maintain muscle function  
109 whilst minimising further tissue damage (24–26). In support of this, recent research using  
110 high-density surface electromyography has revealed that pain causes distinct adaptations  
111 throughout the motor unit pool, including the excitation of higher-threshold units and the  
112 inhibition of lower-threshold units (26). Similarly, the function of the corticospinal pathway  
113 in response to acute muscle pain has also been studied using transcranial magnetic  
114 stimulation (TMS). This produces a motor-evoked potential (MEP) of which the amplitude  
115 can reflect corticospinal excitability, and the duration of the corticospinal silent period during  
116 an active contraction reflects inhibition of the corticospinal pathway. MEP amplitudes have  
117 been shown to increase (25, 27, 28) and decrease (25, 29–31) in response to acute muscle  
118 pain though. Inconsistent responses between studies may be influenced by the muscles tested,

the presence of muscle contraction (i.e., active motor state) or the TMS stimulus intensities used. Recently, Škarabot et al. (32) demonstrated that increasing TMS intensity caused an orderly increase in the recruitment of motor units in the evoked response. Thus, based on this principle, low and high stimulation intensities (relative to the motor threshold) may provide further insight about the corticospinal adjustments for different populations of corticospinal neurons (low and high threshold) in response to acute muscle pain.

Research investigating the aforementioned effects of pain have used a variety of methods, such as blood flow restriction and exercise-induced muscle damage. However, these are commonly conducted contralateral to the muscle of interest, due to the changes in muscle oxygenation (33) or disruption to excitation-contraction coupling process, which preclude the ability to study the effects of *localised* pain on neuromuscular function. One approach is the hypertonic saline pain model, which is primarily used in a sample of healthy participants. This method non-specifically activates group III/IV afferents, which notably elicits an artificial pain response comparable with the experience of natural exercise-induced pain, involves the infusion of a small bolus of hypertonic saline into the muscle (6, 13, 34). An advantage of the hypertonic saline model is that it does not appear to directly affect the contractile properties of muscle fibres (14) and can be procedurally matched with a non-painful isotonic saline injection to serve as a control comparison (6).

Therefore, to gain further insight into the motor performance effects of localised, acute muscle pain on the lower-limb, we utilised the hypertonic saline model of muscle pain to assess RFD at different phases of the contraction along with neuromuscular function (maximal voluntary force, voluntary activation, potentiated twitch force [ $Q_{tw}$ ]). Additionally, to further test the hypothesis that pain has non-uniform inhibitory effects on the neuromuscular system, we investigated a variety of corticospinal responses in the painful vastus lateralis (VL) and a non-painful synergist muscle (rectus femoris [RF]). Specifically,

we explored corticospinal excitability and inhibition across different populations of corticospinal neurons by stimulating the motor cortex at low and high TMS intensities. It was hypothesised that acute muscle pain would reduce maximal voluntary force, RFD, and voluntary activation. Furthermore, during low TMS intensities, it was hypothesised that corticospinal excitability would decrease, and corticospinal inhibition would increase, whereas during high stimulation intensities, corticospinal excitability would increase, and corticospinal inhibition would decrease for both the VL and RF.

## **Methods**

### Participants

Fifteen healthy participants (mean  $\pm$  SD age:  $28 \pm 7$  years; height:  $1.79 \pm 0.08$  m; mass:  $86.9 \pm 16.8$  kg), including 5 females, volunteered to participate in this study. Prior to the commencement of testing, participants completed a physical activity readiness questionnaire and provided written informed consent. A study-specific health questionnaire was completed to screen for contraindications to TMS (35) and intramuscular injections. Participants with neurological disorders, blood-borne diseases, a phobia of needles, any food allergies, lower limb injuries and anyone taking medication for pre-existing pain were excluded from the study; no participants were taking analgesics at the time of the study. Participants were required to abstain from alcohol 24 h, caffeine 4 h, analgesics 6 h and strenuous lower limb physical activity 48 h prior to all testing. This study conformed to the standards of the Declaration of Helsinki (except for pre-registration) and ethical approval was granted by the St Mary's University, Twickenham Ethics Committee (approval reference: SMU\_ETHICS\_2023-24\_460).

### Sample Size Justification

171 The sample size required for the study was calculated a-priori using G\*Power (36).  
172 An effect size of  $dz = 1.02$  from published literature (14) was used which compared the  
173 absolute maximal voluntary force in Newtons measured one minute after a hypertonic versus  
174 isotonic saline injection. To achieve 95% power ( $\beta = 0.95$ ) with an alpha level of 0.05 using a  
175 two-tailed paired samples t-test, a total of fifteen participants were required. Additionally,  
176 given the stark contrast between two interventions (pain vs no-pain) we expected large effect  
177 sizes to be observed. Sensitivity analysis reveals that normal conventions of  $\beta = 0.80$  and  
178 alpha of 0.05 with  $n = 15$ , it was possible to reliably detect Cohen's  $dz = 0.78$ .

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## 180 Experimental Design

181 Following a randomised, crossover design, participants were required to attend the  
182 laboratory on three separate occasions interspaced by 3-8 days. In visit one, participants'  
183 stature and body mass were measured, and they were familiarised with all experimental  
184 procedures, including an intramuscular injection of hypertonic saline, along with pain related  
185 perceptual measures and questionnaires (see Perceptual Measures). The following two  
186 experimental trials were completed in a randomised, incomplete-counterbalanced order.  
187 These included baseline neuromuscular function tests (electromyography [EMG], maximal  
188 compound muscle action potential [ $M_{\max}$ ], maximal voluntary contractions [MVC] and TMS)  
189 prior to performing the experimental procedure. Pre-injection, participants performed one 4-5  
190 s MVC to assess maximal voluntary force and RFD with a superimposed and resting  
191 peripheral nerve stimulation delivered to the femoral nerve to assess quadriceps voluntary  
192 activation and  $Q_{\text{tw}}$ . Subsequently, three sets of seven intermittent contractions at 20% of the  
193 MVC force determined at baseline were performed ( $\sim 3$  s on,  $\sim 3$  s off) with TMS delivered  
194 at 120%, 150% and 80/120% active motor threshold (AMT) (paired pulse, 3 ms inter-  
195 stimulus interval) to quantify corticospinal excitability, inhibition and short interval



intracortical inhibition (SICI). The order of the sets was assigned randomly to each participant and counterbalanced (i.e., five participants had 120% AMT first, five had 120% second and five had 120% third), but the order was kept consistent across trials for the same participant. A final resting peripheral nerve stimulation was delivered after the TMS to normalise corticospinal responses to. Approximately five minutes after completion of the pre-injection procedures, participants received an intramuscular injection of either isotonic saline (ISO) or hypertonic (HYP) into the vastus lateralis. After the injection, participants were seated back in the isometric chair and completed the post-injection procedures, which were identical to pre-injection. Temporally, the time taken from needle removal to commencement of the post-injection procedures was approximately 60 s and the post-injection procedure took approximately 150 seconds. After the post-injection procedures were completed, participants completed the long-form McGill pain questionnaire. A schematic of the experimental visits can be seen in Figure 1.

## Equipment and Procedures

*Experimental Muscle Pain.* To induce acute muscle pain, participants received an intramuscular injection of hypertonic saline while seated at rest on the edge of a medical bed in a wet laboratory < 5 m from the isometric chair. In the familiarisation session a single bolus of the hypertonic saline solution (1 mL, 5.85% [B Braun Medical Industries]) was injected into the vastus lateralis (VL) of the right leg (middle third of the lateral head of the thigh between the lateral femoral condyle of the femur and the greater trochanter) while the participant's knee was at a 90° angle (34). In the experimental visits, participants received either the hypertonic saline or isotonic saline (1 mL 0.9%) injection using a 25-gauge, 25 mm, Luer-Lok, hypodermic needle (BD Microlance, Switzerland), connected to a 5 mL syringe (BD Microlance, Switzerland). Both the main researcher and participant were blinded

to the injection until administration. The injection site and surrounding area were palpated and inspected to identify no local tenderness/muscle soreness before injections. All injections were administered using the Z-track method (37). Injections were performed manually for 20 seconds (5 s pause following needle insertion and aspiration, 10 s of solution infusion [infusion rate  $\sim 0.1$  mL/s] and 5 s pause before need removal). An aspiration was performed after the needle insertion to confirm the needle was not in a blood vessel (14). Immediately after needle withdrawal, participants moved onto the isometric chair.

*Mechanical Recordings.* Participants were strapped into a custom-built isometric chair with a knee and hip angle of  $90^\circ$ . A Velcro strap was fastened 2 cm above the right malleoli. The strap was connected to a linear force transducer (FSB- 1.5 kN Universal Cell 1.5 kN, Force Logic, Reading, UK) to measure knee extensor isometric force. A data capture module (CED Micro 1401, CED, Cambridge, UK) sampled force data onto compatible software (Signal V8, CED, Cambridge, UK), at a frequency of 2.5 KHz. Instantaneous feedback of the force traces was provided to participants on a screen directly in front of them.

*Maximal Voluntary Contractions.* Participants were instructed to contract ‘as hard and as fast as possible’ for 4 – 5 seconds. Three MVCs  $\sim 2$  minutes apart were carried out at baseline to confirm participants were familiarised with MVCs, and to determine the 20% MVC force required for the subsequent TMS measurements. During the pre- and post-injection protocol, participants were required to do a single MVC. To assess voluntary activation and  $Q_{tw}$ , a peripheral nerve stimulation was delivered at peak force of the MVC and at rest (38).

*Peripheral Nerve Stimulation.* Electrical stimulations were delivered to the femoral nerve to innervate the right quadriceps femoris using an electrical stimulator (DS7AH constant-current stimulator, Digitimer, Welwyn Garden City, UK) (maximum voltage of 400 V) that delivered a single square wave pulse (200  $\mu$ s duration). Two self-adhesive 32  $\times$  32 mm circular self-adhesive neurostimulation electrodes (Axelgaard Manufacturing, Lystrup, Denmark) were placed on the right gluteal fold (anode) and within the femoral triangle (cathode). The correct placement of the cathode was confirmed when an observable twitch response was achieved at a stimulation intensity of 100 mA. Electrical stimuli were then delivered in 20 mA increments starting from 60 – 100 mA depending on initial response to identify the stimulation intensity that resulted in a plateau in the M-wave peak-to-peak amplitude. An additional 20% of the stimulation intensity was delivered to ensure a supramaximal stimulus was delivered (39).

*Electromyography.* Muscle activity of the VL and RF was recorded using 36 mm  $\times$  36 mm bipolar surface electrodes (WhiteSensor 4500M, Ambu Ltd, Denmark) with a 20 mm interelectrode distance. Electrode sites were identified using the SENIAM guidelines and a reference electrode was placed on the right patella. The electrode sites were shaved, abraded, and cleaned with an alcohol swab to improve conductivity (40). All EMG signals were recorded at 2.5 kHz and amplified (gain 1000) using a signal amplifier (D440-2-Two Channel Isolated Amplifier, Digitimer, Welwyn Garden City, UK) before being recorded onto compatible software and bandpass filtered (10-1000 Hz) (Signal V8, CED, Cambridge, UK).

*Transcranial Magnetic Stimulation.* Using a 110 mm double-cone coil, single-pulse and paired-pulse TMS was delivered to the motor cortex with two magnetic stimulators (Magstim Bistim, The Magstim Company Ltd, Whitland, UK). The procedure started with

determining the hotspot by marking on the participant, who was wearing a skintight Lycra swimming hat, the vertex, which was measured as the midpoint between the tragus and nasalinion. TMS pulses at 35-45% maximal stimulator output were delivered during a submaximal isometric contraction (20% MVC) at 0, 1 and 2 cm anteriorly and posteriorly from the marked vertex. The anterior/posterior location which evoked the greatest MEP response was then further marked with 1 cm and 2 cm marks to the left. The greatest MEP peak-to-peak amplitude in the vastus lateralis out of all of these stimulations was defined as the hotspot. Following this, five stimulations were delivered at 45 – 55% maximal stimulator output during a 20% MVC contraction and the stimulator intensity was increased or decreased in increments of 1 or 5% until the lowest intensity in which three out of the five stimulations were > 0.2 mV and had a visible silent period was reached. This was defined as the participant's active motor threshold (AMT). Subsequent stimulations at pre- and post-injection were set at 120% and 150% for single-pulse and 80/120% AMT for paired-pulse with an inter stimulus interval of 3 ms. A stimulation intensity of 150% was selected to assess the behaviour of higher-threshold corticospinal neurons in comparison to low-threshold that are commonly recruited during a 120% stimulation intensity (32).

*Perceptual Measures.* A custom-built, electronic visual analogue scale (VAS) was used to record pain intensity (41) during the post-injection procedures. Participants began rating their pain once seated back in the isometric chair. The scale ranged from 0 ('no pain at all') to 100 ('Extremely intense pain [almost unbearable]'). Participants were instructed to rate their pain in relation to the worst exercise-induced pain they have previously experienced and *not* based on previous injuries or their worst imaginable pain. Participants adjusted the slider on the scale accordingly (see figure 2). After the post-injection procedures, the long-form McGill pain questionnaire was administered (42) to determine the quality of pain

experienced following the injection. Participants were explicitly instructed not to include any pain from the needle stick or electrical/magnetic stimulations.

### Data Analysis

All data analysis were performed in a blinded design to minimise experimenter bias. Blinding of data files was achieved by having a separate researcher code the data files to a random number that did not correspond to the trial the participant had completed for that session. This was only revealed after data analysis was complete.

RFD was calculated from each MVC as the slope of the first 200 ms of the rising force-time curve, separated into 50 ms epochs. Contraction onset was visually determined as the last trough of the resting force trace before a rise above baseline. Force traces were observed on a consistent X (1 s) and Y axis scale ( $\pm 37.5$  N around the resting force trace) on a 24.5-inch computer monitor (Acer Nitro XF252Q, Acer, New Tapei City, Taiwan).

Voluntary activation was calculated with the following equation (38):

$$100 - \text{superimposed twitch force (N)} \frac{\left( \frac{\text{force before twitch (N)}}{\text{peak force (N)}} \right)}{\text{resting potentiated twitch force (N)}} \times 100$$

Corticospinal excitability was determined as the average of the peak-to-peak amplitudes of the MEPs, which were then normalised to the peak-to-peak amplitude of the  $M_{\max}$  ( $\text{MEP} \cdot M_{\max}$ ). The duration of the TMS silent period was visually inspected from the point of the stimulus artefact to the resumption of voluntary EMG activity; The average of these durations reflected corticospinal inhibition. SICI was calculated as:

$$\left( 1 - \frac{\text{mean of conditioned MEPs (mV)}}{\text{mean of unconditioned MEPs (mV)}} \right) \times 100$$

If a lower number was observed, this reflected less inhibition, and vice versa (43). Analysis of the McGill pain questionnaire was separated into the components set out in the form. These components categorise pain into sensory (boxes 1 – 10), affective (boxes 11-15),

evaluative (box 16) and miscellaneous (boxes 17 – 20). The pain intensity selected for analysis was the single value recorded by the VAS the moment before each MVC and set of stimulations was performed. A basic frequency analysis was performed to determine commonly chosen words across the sample, which was set as any word selected by more than one third of participants.

### Statistical Analysis

All statistical analyses were conducted in JAMOV 2.5.3 (The Jamovi Project, 2024). The intensity of pain reported immediately prior to the MVC, TMS at 120, 150% AMT, and SICI were analysed with a paired samples t-test. To confirm pain ratings were not different during each of these, a one-way repeated measures analysis of variance (ANOVA) was used to compare pain intensities in HYP only. The  $\Delta$  maximal voluntary force,  $Q_{tw}$  and voluntary activation from pre to post injection for ISO and HYP were analysed with a two-tailed paired samples t-test. Data which did not reasonably meet the assumption of normality (voluntary activation) were analysed with a Wilcoxon sign rank test.

RFD and TMS variables were analysed with a repeated measures linear mixed effects model using the ‘gamlj’ package in JAMOV. For RFD, contraction time (50, 100, 150 and 200 ms) and condition (ISO and HYP) were included as fixed effects. For the TMS silent period and MEP·Mmax, stimulation intensity (120% and 150% AMT), muscle (VL and RF) and condition were included as fixed effects. For SICI, only condition and muscle were included as fixed effects. Individual participant intercepts were included as a random effect. The pre-injection values were included as a covariate to account for any between-session variability of the dependent variables, and for TMS variables, the MVC force which preceded TMS was also included as a covariate to account for any changes in the relative contraction strength which may influence corticospinal excitability/inhibition (44). Once models were

fitted, normality of residuals were assessed using the Kolmogorov-Smirnov test, along with histograms and Q-Q plots. Absence of heteroscedasticity was verified by visually observing the residuals scatterplot. Variables which demonstrated heteroscedasticity (RFD) were log<sup>10</sup> transformed but presented in their original scale for ease of interpretation. A simple effects analysis was performed to determine differences in each factor at each level when a statistically significant interaction effect was observed.

As subsequent exploratory analyses, we investigated whether between pain intensity associated with changes in key outcome variables. Pearson correlations were performed on the  $\Delta\%$  MVC,  $\Delta$  VA and  $\Delta\%$  RFD at 50 ms, as well as  $\Delta$  MEP·Mmax,  $\Delta$  TMS SP and  $\Delta$  SICI measures. Correlation *P*-values were corrected for multiplicity using a Holm-Bonferroni correction. Statistical significance was set at  $P < 0.05$ . Cohen's *d* effect sizes were reported with values of 0.2, 0.5 and 0.8 representing thresholds for small, medium, and large effects respectively (Fritz et al., 2012).

## Results

### VAS (Pain Intensity)

There was a greater rating of pain during the MVC in HYP ( $31 \pm 11$  mm) compared to ISO ( $3 \pm 5$  mm) (mean difference = 28 mm,  $t_{14} = 9.33$ ,  $P < 0.001$ , ES = 2.41). During the 120% AMT TMS, there was also greater pain ratings in HYP ( $33 \pm 16$  mm) than ISO ( $0 \pm 1$  mm) (mean difference = 33 mm,  $t_{14} = 7.96$ ,  $P < 0.001$ , ES = 2.05) as well as during the 150% AMT TMS (HYP =  $37 \pm 19$  mm) compared to ISO ( $0 \pm 1$  mm) (mean difference = 37 mm,  $t_{14} = 7.68$ ,  $P < 0.001$ , ES = 1.98). Lastly, during SICI, there was a significantly higher rating of pain in HYP ( $33 \pm 18$  mm) compared to the ISO ( $0 \pm 0$  mm) (mean difference = 33 mm,  $t_{14} = 6.93$ ,  $P < 0.001$ , ES = 1.79). There was no difference in pain intensity reported during the

367 MVC, or any of the TMS measures in HYP ( $F_{3,42} = 1.98$ ,  $P = 0.132$ ). Individual pain intensity  
 368 ratings are presented in Figure 3.

369

370 McGill long form pain questionnaire

	ISO	HYP
Sensory	-	Cramping (67%) Throbbing (47%) Aching (33%) Sharp (33%) Tender (33%)
SRI	0 [0 – 0.5]	12 [9 – 16]*

371 Wilcoxon sign rank tests revealed a greater subclass rating index in HYP compared to  
 372 ISO for sensory (Wilcoxon  $P < 0.001$ ), evaluative (Wilcoxon  $P = 0.014$ ) and miscellaneous  
 373 (Wilcoxon  $P = 0.003$ ) subclasses, but not for the affective component (Wilcoxon  $P = 0.371$ ).  
 374 Table 1 shows the median and interquartile ranges for subclass rating index values, along  
 375 with most commonly selected words by participants.

376 **Table 1.** McGill long form pain questionnaire most commonly selected words and subclass  
 377 rating index (SRI). Data presented as median and interquartile range.



Affective	-	-
SRI	0 [0 – 0]	0 [0 – 0]
Evaluative	-	-
SRI	0 [0 – 0]	1 [0 – 3]*
Misc.	-	-
SRI	0 [0 – 0]	2 [1 – 4]*

\*denotes significantly different from ISO (Wilcoxon  $P < 0.05$ ).

### Neuromuscular Function

*Maximal Voluntary Force and RFD.* There was a greater decrease of MVC force in HYP compared to the ISO (mean difference = -73 N,  $P = 0.020$ ,  $ES = -0.68$ ). For RFD, a condition  $\times$  contraction phase interaction was observed ( $F_{3, 96.4} = 2.76$ ,  $P = 0.047$ ). Simple effects analysis revealed RFD was lower in HYP compared to ISO at 50 ms (mean difference =  $-313 \text{ N}\cdot\text{s}^{-1}$ ,  $P = 0.029$ ,  $ES = -0.57$ ), 100 ms (mean difference =  $-617 \text{ N}\cdot\text{s}^{-1}$ ,  $P = 0.010$ ,  $ES = -0.68$ ) and 150 ms (mean difference =  $-525 \text{ N}\cdot\text{s}^{-1}$ ,  $P = 0.013$ ,  $ES = -0.66$ ) of contraction onset, but not at 200 ms (mean difference =  $214 \text{ N}\cdot\text{s}^{-1}$ ,  $P = 0.406$ ,  $ES = 0.22$ )(Figure 4).

*Voluntary Activation,  $Q_{tw}$  and M-Wave Amplitude.* There was a greater decrease of VA in HYP compared to ISO (median difference = -1.3%,  $P = 0.008$ ,  $ES = -0.66$ ). No difference in  $\Delta Q_{tw}$  between HYP and ISO was observed (mean difference = -11 N,  $P = 0.066$ ,  $ES = -0.51$ ) (figure 5). Because  $Q_{tw}$  revealed moderate effect sizes after the MVC, we did exploratory analysis on the  $Q_{tw}$  obtained at the end of the trial (i.e., after all TMS was completed) which revealed no significant differences from ISO to HYP (median difference = -0.8 N,  $P = 0.934$ ,  $ES = 0.03$ ). M-Wave amplitude of the VL (injected muscle) was also not different between ISO and HYP (mean difference = -0.04 mV,  $P = 0.844$ ,  $ES = -0.05$ ).

### TMS Responses

399 *Corticospinal Excitability (MEP·M<sub>max</sub>)*. There was no condition × muscle × intensity  
 400 ( $F_{1,95.9} = 0.050$ ,  $P = 0.824$ ), condition × intensity ( $F_{1,96.0} = 0.539$ ,  $P = 0.465$ ) or condition ×  
 401 muscle ( $F_{1,92.2} = 0.780$ ,  $P = 0.379$ ) interaction. There was a significant fixed effect of  
 402 condition ( $F_{1,107} = 3.354$ ,  $P = 0.023$ ) with MEP·M<sub>max</sub> being greater in HYP compared to ISO  
 403 (mean difference = 4%,  $P = 0.023$ , ES = 0.60). Significant fixed effects of intensity ( $F_{1,108.6} =$   
 404 4.686,  $P = 0.033$ ) revealed MEP·M<sub>max</sub> was greater in 150% AMT compared to 120% AMT  
 405 (mean difference = 4%,  $P = 0.033$ , ES = 0.56). No effect of muscle was observed ( $F_{1,108.6} =$   
 406 0.68,  $P = 0.413$ ).

407 *Corticospinal Inhibition (TMS Silent Period Duration)*. No condition × muscle ×  
 408 intensity ( $F_{1,97.3} = 0.005$ ,  $P = 0.943$ ) or condition × muscle ( $F_{1,97.3} = 0.231$ ,  $P = 0.632$ )  
 409 interaction effects were observed, but there was a significant condition × intensity interaction  
 410 ( $F_{1,98.6} = 6.294$ ,  $P = 0.014$ ). Simple effects revealed that silent period was not different  
 411 between ISO and HYP at 120% AMT (mean difference = 3.1 ms,  $P = 0.280$ , ES = 0.28),  
 412 however at 150% AMT silent period duration was shorter in HYP compared to ISO (mean  
 413 difference = -7.0 ms,  $P = 0.017$ , ES = 0.63). A significant fixed effect for intensity was  
 414 observed ( $F_{1,108.2} = 14.576$ ,  $P < 0.001$ ), with silent periods being longer at 150% AMT  
 415 compared to 120% (mean difference = 9.1 ms,  $P < 0.001$ , ES = 0.99).

416 *SICI*. There was no condition × muscle interaction ( $F_{1,41.8} = 0.036$ ,  $P = 0.851$ ), but  
 417 there was a fixed effect of condition ( $F_{1,46.1} = 4.587$ ,  $P = 0.038$ ) where SICI was greater in  
 418 HYP compared to ISO (mean difference = 8.3%,  $P = 0.038$ , ES = 0.55).

419 **Figure 6.** TMS responses between ISO and HYP post-injection. a. MEP·M<sub>max</sub> indicative of  
 420 corticospinal excitability. b. TMS Silent period duration, indicative of corticospinal  
 421 inhibition. c. Short interval intracortical inhibition, indicative of cortical inhibition. \* denotes  
 422 significantly different from ISO in 150% AMT in both VL and RF muscles ( $P < 0.05$ ). Data

presented as estimated marginal mean  $\pm$  95% confidence interval. Only significant fixed effects are presented alongside figures.

## Correlations

There was no significant relationship between pain intensity and any neuromuscular outcome (all  $P \geq 0.228$ ). Specific correlations,  $r$ , and  $P$  values can be observed in supplementary ‘correlations’ analysis file.

## **Discussion**

The aim of the present study was to assess neuromuscular function and corticospinal responses to acute muscle pain induced by an injection of hypertonic saline into the VL. The principle novel findings are as follows i.) Muscle pain caused a significant reduction in RFD during the initial phase of the MVC. ii.) Muscle pain induced both excitatory and inhibitory responses, with an increase in MEP·M<sub>max</sub> and SICI, whereas corticospinal silent period duration displayed reduced inhibition at higher stimulation intensities but was unaffected at low stimulation intensities. Furthermore, our data supports previous observations (6, 13) that muscle pain reduces maximal force-generating capacity, which appears to be due to central and not peripheral mechanisms.

## Perceptions of Pain Induced by Hypertonic Saline

An intramuscular injection of hypertonic saline into the vastus lateralis induced, on average, moderate (30-37/100) intensities of muscle pain (figure 3), which was sustained at this intensity for the entire duration of the post-injection procedures. These pain intensities are somewhat lower what has previously been observed (45-60/100) in response to 1 mL of 5.85% NaCl injected into the vastus lateralis (16, 23, 34). One explanation for this lower pain response could be the presence of exercise-induced hypoalgesia caused by MVCs prior to the

injections (45). In line with previous research, our results from the McGill questionnaire demonstrated a high sensory component (table 1) from the hypertonic saline injection with commonly described words such as “aching” and “throbbing” (16, 34, 46). Affective and evaluative components were low, which was expected given the familiar, transient, and non-damaging nature of the experimental pain model.

#### Effects of Pain on Neuromuscular Function

Elevated muscle pain from the hypertonic saline injection caused a ~10% (large effect size) reduction in knee-extensor MVC force (figure 5a), which in agreement with the previous literature which has consistently observed a 7.5 – 21% lower MVC force after an intramuscular hypertonic saline injection compared to an isotonic saline injection (12–15). In consonance with a lower MVC force in HYP was a lower voluntary activation (figure 5b), indicating that the impairment in maximal force production was due to central fatigue. It is plausible that the reduced voluntary activation is caused by a conscious disengagement from the task (i.e., reduced effort), rather than central fatigue. However, given that there were low affective and evaluative components of the pain response, it would be unlikely that the participant would have a strong reason to voluntarily apply less effort. Additionally, our experimental model of pain was tonic (as opposed to movement evoked), thus the performance of the MVC would have had no immediate negative effect on the pain experience. Voluntary disengagement is likely present during movement evoked pain, compared to tonic pain (47). Therefore, we contend that the reduced voluntary activation observed in the present study is due to a suboptimal central drive from the motor cortex.

There was no significant difference in  $Q_{tw}$  or M-Wave amplitude between trials. This further supports the possible central mechanistic involvement and absence of peripheral mechanisms during pain evoked by a hypertonic saline injection. Whilst a moderate effect

size was observed for a lower  $Q_{tw}$  in HYP, this may be due to a small loss of potentiation from a lower absolute MVC force induced by pain (48). Indeed, when evaluating the  $Q_{tw}$  at the end of the testing procedures where potentiation would be more consistent between trials, the effect size was trivial. Taken together, these findings support the wider literature to show that acute muscle pain induced by an intramuscular injection of hypertonic saline does not directly affect the peripheral fatigue (14, 49, 50).

This is the first study to show that RFD was impaired by a moderate degree ( $ES = -0.57$  to  $-0.68$ ) in the presence of experimentally induced muscle pain, but only for the first 150 ms of the contraction (Figure 4). It been established that the early and late components of RFD are limited by different physiological factors (51). For example, during the early RFD phase ( $< 75$  ms), maximum motor unit discharge rate is crucial for maximising the initial force production, whereas the muscle's contractile properties have a greater influence on the latter component (52). As mentioned previously, hypertonic saline-induced acute muscle pain has no influence on the contractile properties of a muscle, which could explain why the late phase (150-200 ms) was unaffected by pain. Similar findings have been observed by Rice et al. (2019), but with experimental knee pain induced by hypertonic saline injections into the infrapatellar fat pad. Motor unit discharge rates have been shown to decrease in lower threshold motor units and increase in higher threshold motor units in response to experimental muscle pain. However, it appears that this acute compensatory process was unable to preserve early-phase RFD, which presumably relies upon maximal discharge rates being achieved across the entire motor unit pool (54).

#### Effect of Pain on Corticospinal Responses

Corticospinal adjustments induced by hypertonic saline were assessed during low-force voluntary contractions. We found an increase in corticospinal excitability following the

hypertonic saline injection (figure 6a); which was not dependent on the stimulation intensity used, crudely suggesting increased excitability across much of the corticospinal neuronal pool, and potentially, both low and higher threshold motor units. When assessed at rest, pain has generally been shown to cause a reduction in corticospinal excitability (55). Previous work, however, has reported an increase in the lower limb corticospinal excitability when assessed at rest during experimental knee pain (56, 57). To our knowledge, assessment of corticospinal excitability during active contractions in response experimental lower-limb muscle pain has only previously been studied once (14). They found no differences in VL MEP amplitudes after a 1 mL hypertonic saline injection into the VL. However, it should be noted that these measurements were recorded during a fatiguing isometric knee extensor task which may introduce corticospinal adjustments in response to exercise (58) rather than solely pain. Therefore, our results are the first to demonstrate an increase in corticospinal excitability of the VL and RF in response to quadriceps acute muscle pain. Alongside increased excitation, there was evidence of reduced corticospinal inhibition during HYP which was reflected by a shorter corticospinal silent period at 150% AMT (figure 6b). Interestingly, no such difference was observed for stimulations at 120% AMT. Research by Martinez-Valdes et al. (26) found that lower-threshold motor units were able to maintain their discharge rate at both low and high contraction intensities (0 – 70% MVC) compared to the higher-threshold motor units that were shown to increase discharge rate and lower their recruitment threshold. These low threshold motor units are more susceptible to inhibitory input and are more affected by persistent inward currents (59). However, high-threshold units are not largely dependent on persistent inward currents but on increased excitatory input (60). Therefore, it is possible that our demonstration of reduced corticospinal inhibition at higher stimulation intensities was reflective of the behaviour of higher-threshold motor units, lending support to the notion that the decreased inhibition during HYP may be associated

with these motor units increasing their discharge rate and lowering their recruitment threshold (26). Paired-pulse TMS measuring SICI, however, revealed an increase in inhibition (figure 6c). Given that these two measures reflect different inhibitory mechanisms, (SICI; GABA<sub>A</sub> and TMS silent period duration; GABA<sub>B</sub>) it is plausible that the balance of these are partly responsible for regulating motor output during pain. Whilst the role of each specific corticospinal adjustment is not fully understood, it has been proposed that at the system level, these adjustments allow for motor tasks to be maintained, whilst minimising further pain or tissue damage (24). However, a consequence of these neurophysiological changes is a reduced maximal force-generating capacity, decreased endurance capacity and greater perceived effort (14, 61).

### Physiological Relevance

It is important to note that whilst hypertonic saline induces sensations of pain, due to depolarisation of both group III/IV nociceptors (62), there may be activation of other non-nociceptive afferents with this technique. This is due to the relatively non-specific stimulation of free-nerve endings induced by increasing extracellular sodium concentrations (63). However, the relative contribution of these non-nociceptive afferents is argued to be minor (64). Therefore, our data provide insight into the neuromuscular responses following generalised nociceptive and to a lesser-degree, non-nociceptive stimulation. Interestingly, recent evidence has shown that the neuromuscular responses to group III/IV afferent stimulation may be dependent on the specific sub-types involved (e.g., nociceptive vs non-nociceptive afferents) (65).

Given that isometric contractions themselves are sufficient to activate mechanoreceptor afferents, any additional effect from hypertonic saline on these afferents is likely minimal. Supporting this, recent work by Zambolin et al. (65) demonstrated that

neuromuscular responses to mechano-nociceptive stimulation were generally directionally similar to those evoked by non-painful mechanical stretch, with greater afferent loading—primarily driven by nociceptive input—modulating the magnitude of response. This highlights that the hypertonic saline model likely exerts its primary effect through the addition of nociceptive afferents, rather than novel mechanoreceptor recruitment.

While recent studies have employed models such as blood flow restriction (BFR) and exercise-induced muscle damage (EIMD) to explore afferent activation and neuromuscular responses, these approaches differ in key respects from hypertonic saline infusion. BFR induces systemic cardiovascular changes and discomfort that may confound localised nociceptive input (33, 66), while EIMD triggers a cascade of secondary processes, including inflammation and structural muscle changes (67), that complicate the isolation of pain-related effects. In contrast, hypertonic saline provides a well-established, controllable model for inducing localised, tonic muscle pain without the accompanying metabolic or mechanical stimuli present in other paradigms (66). This allows for more precise investigation of the localised effects of nociceptive afferent activity on neuromuscular function, particularly during isometric tasks. Despite being a less “naturalistic” pain model, its internal validity and reproducibility make it highly relevant for mechanistic studies of pain and motor control.

#### Methodological Considerations

The inter-individual variability in pain responses from participants in the present study (figure 3) may have resulted in heterogenous outcomes between participants. In particular, low levels of pain reported by some participants likely introduced some uncertainty around the findings. It was not feasible to exclude these participants as this would have significantly compromised statistical power, but these findings may be limited in terms



of their application to a specific pain intensity. Future studies could implement an individualised pain induction approach by varying injection volumes to evoke a desired pain intensity (e.g., 'mild,' 'moderate' or 'Strong' on the VAS). Alternatively, studies may want to assess motor performance across the descending limb of the pain response to better quantify the impact of high and low pain intensities on motor performance.

Due to the relatively short time frame of hypertonic saline evoked pain, we were limited our protocol to 21 TMS pulses (7 pulses for 3 different stimulation parameters). Previous work has indicated that >18 pulses may be needed to obtain a 'true' measure of corticospinal excitability or SICI (68). Therefore, our relatively low number of stimulations may introduce some additional variability. However, even with 5 stimulations, in this study we achieved within-session ICC point estimates at >0.80 which is considered 'good' reliability (68, 69). Future research may want to explore additional neurophysiological measures, such as intracortical facilitation, long-interval intracortical inhibition or even spinal excitability to gain further insight into the effects of acute quadriceps muscle pain.

Five females participated in this study; however, the phases of the menstrual cycle were not controlled for. Neuromuscular function has been suggested to be reduced during the follicular phase (64); however, more recent research has found during gross motor movements, there is no change in strength and neuromuscular function for both the early and late follicular phases (71).

## **Conclusion**

In summary, experimentally induced muscle pain in the quadriceps reduced knee extensor MVC force, RFD and voluntary activation compared to a non-painful isotonic saline injection. Pain exerts both excitatory and inhibitory effects on the corticospinal pathway. These findings provide further evidence to support the notion that the neuromuscular system

can maintain task demands during submaximal contractions, albeit with altered excitability and inhibition, but as a consequence maximal motor task performance is impaired. These findings have implications for a wide range of individuals who experience muscle pain in the knee extensors whilst performing maximal and submaximal motor tasks.

## **Additional Information**

Data Availability Statement: The data used for statistical analysis and individual data analysis files can be found at the following link: <https://doi.org/10.6084/m9.figshare.28271465.v1>

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624 investigated and resolved; and all persons designated as authors qualify for authorship, and  
625 all those who qualify for authorship are listed.

## References

1. **Raja SN, Carr DB, Cohen M, Finnerup NB, Flor H, Gibson S, Keefe FJ, Mogil JS, Ringkamp M, Sluka KA, Song XJ, Stevens B, Sullivan MD, Tutelman PR, Ushida T, Vader K.** The Revised IASP definition of pain: concepts, challenges, and compromises. *Pain* 161: 1976, 2020. doi: 10.1097/J.PAIN.0000000000001939.
2. **Bruehl S.** Complex regional pain syndrome. *BMJ* 351, 2015. doi: 10.1136/BMJ.H2730.
3. **Staud R.** Biology and therapy of fibromyalgia: Pain in fibromyalgia syndrome. *Arthritis Res Ther* 8: 1–7, 2006. doi: 10.1186/AR1950/FIGURES/2.
4. **Zambolin F, Duro-Ocana P, Faisal A, Bagley L, Gregory WJ, Jones AW, McPhee JS.** Fibromyalgia and Chronic Fatigue Syndromes: A systematic review and meta-analysis of cardiorespiratory fitness and neuromuscular function compared with healthy individuals. *PLoS One* 17: e0276009, 2022. doi: 10.1371/JOURNAL.PONE.0276009.
5. **Franklin JD, Atkinson G, Atkinson JM, Batterham AM.** Peak Oxygen Uptake in Chronic Fatigue Syndrome/Myalgic Encephalomyelitis: A Meta-Analysis. *Int J Sports Med* 40: 77–87, 2019. doi: 10.1055/A-0802-9175/ID/R7132-0017/BIB.
6. **O'Malley CA, Smith SA, Mauger AR, Norbury R.** Exercise-induced pain within endurance exercise settings: Definitions, measurement, mechanisms and potential interventions. .
7. **Cheung K, Hume PA, Maxwell L.** Delayed onset muscle soreness: Treatment strategies and performance factors. *Sports Medicine* 33: 145–164, 2003. doi: 10.2165/00007256-200333020-00005/METRICS.
8. **Iannetta D, Zhang J, Murias JM, Aboodarda SJ.** Neuromuscular and perceptual mechanisms of fatigue accompanying task failure in response to moderate-, heavy-, severe-, and extreme-intensity cycling. *J Appl Physiol* 133: 323–334, 2022. doi: 10.1152/JAPPLPHYSIOL.00764.2021/ASSET/IMAGES/LARGE/JAPPLPHYSIOL.00764.2021\_F004.JPEG.
9. **Spitz RW, Wong V, Bell ZW, Viana RB, Chatakondi RN, Abe T, Loenneke JP.** Blood Flow Restricted Exercise and Discomfort: A Review. *J Strength Cond Res* 36: 871–879, 2022. doi: 10.1519/JSC.0000000000003525.
10. **Wender CLA, McGranahan MJ, O'Connor PJ.** Exercise-induced quadriceps pain during cycling in healthy individuals: A systematic review and meta-analysis of experimental trials. *Sci Sports* 38: 827–835, 2023. doi: 10.1016/J.SCISPO.2022.12.005.
11. **Martin PG, Weerakkody N, Gandevia SC, Taylor JL.** Group III and IV muscle afferents differentially affect the motor cortex and motoneurons in humans. *J Physiol* 586: 1277–1289, 2008. doi: 10.1113/jphysiol.2007.140426.
12. **Graven-Nielsen T, Svensson P, Arendt-Nielsen L.** Effects of experimental muscle pain on muscle activity and co-ordination during static and dynamic motor function. *Electroencephalography and Clinical Neurophysiology/Electromyography and Motor Control* 105: 156–164, 1997. doi: 10.1016/S0924-980X(96)96554-6.
13. **Graven-Nielsen T, Lund H, Arendt-Nielsen L, Danneskiold-Samsøe B, Bliddal H.** Inhibition of maximal voluntary contraction force by experimental muscle pain: A centrally mediated mechanism. *Muscle Nerve* 26: 708–712, 2002. doi: 10.1002/MUS.10225.

14. **Norbury R, Smith SA, Burnley M, Judge M, Mauger AR.** The effect of elevated muscle pain on neuromuscular fatigue during exercise. *Eur J Appl Physiol* 122: 113–126, 2022. doi: 10.1007/S00421-021-04814-1/FIGURES/5.
15. **Becker K, Goethel M, Fonseca P, Vilas-Boas JP, Ervilha U.** The Strategy of the Brain to Maintain the Force Production in Painful Contractions—A Motor Units Pool Reorganization. *Cells* 2022, Vol 11, Page 3299 11: 3299, 2022. doi: 10.3390/CELLS11203299.
16. **Smith SA, Micklewright D, Winter SL, Mauger AR.** Muscle pain induced by hypertonic saline in the knee extensors decreases single-limb isometric time to task failure. *Eur J Appl Physiol* 120: 2047–2058, 2020. doi: 10.1007/S00421-020-04425-2/FIGURES/3.
17. **Canestri R, Franco-Alvarenga PE, Brietzke C, Vinícius Í, Smith SA, Mauger AR, Goethel MF, Pires FO.** Effects of experimentally induced muscle pain on endurance performance: A proof-of-concept study assessing neurophysiological and perceptual responses. *Psychophysiology* 58: e13810, 2021. doi: 10.1111/PSYP.13810.
18. **Zhang J, Abel S, Macphail M, Aboodarda SJ.** Persistent Contralateral Pain Compromises Exercise Tolerance but Does not Alter Corticomotor Responses During Repeated, Submaximal Isometric Knee Extensions to Task Failure. *Neuroscience* 526: 267–276, 2023. doi: 10.1016/J.NEUROSCIENCE.2023.07.005.
19. **Aboodarda SJ, Iannetta D, Emami N, Varesco G, Murias JM, Millet GY.** Effects of pre-induced fatigue vs. concurrent pain on exercise tolerance, neuromuscular performance and corticospinal responses of locomotor muscles. *J Physiol* 598: 285–302, 2020. doi: 10.1113/JP278943.
20. **Maffiuletti NA.** Physiological and methodological considerations for the use of neuromuscular electrical stimulation. *Eur J Appl Physiol* 110: 223–234, 2010. doi: 10.1007/S00421-010-1502-Y/METRICS.
21. **Prevc P, Misotic N, Stirn I, Tomazin K.** Perceived Discomfort and Voluntary Activation of Quadriceps Muscle Assessed with Interpolated Paired or Triple Electrical Stimuli. *International Journal of Environmental Research and Public Health* 2023, Vol 20, Page 4799 20: 4799, 2023. doi: 10.3390/IJERPH20064799.
22. **Finn H, Rouffet DM, Kennedy DS, Green S, Taylor JL.** Motoneuron excitability of the quadriceps decreases during a fatiguing submaximal isometric contraction. *J Appl Physiol* 124: 970–979, 2018. doi: 10.1152/JAPPLPHYSIOL.00739.2017/ASSET/IMAGES/LARGE/ZDG0031825550005.JPEG.
23. **Norbury R, Smith SA, Burnley M, Judge M, Mauger AR.** The effect of hypertonic saline evoked muscle pain on neurophysiological changes and exercise performance in the contralateral limb. *Exp Brain Res* 240: 1423–1434, 2022. doi: 10.1007/S00221-022-06342-6/FIGURES/5.
24. **Hodges PW, Tucker K.** Moving differently in pain: A new theory to explain the adaptation to pain. *Pain* 152: S90–S98, 2011. doi: 10.1016/J.PAIN.2010.10.020.
25. **Martin PG, Weerakkody N, Gandevia SC, Taylor JL.** Group III and IV muscle afferents differentially affect the motor cortex and motoneurons in humans. *J Physiol* 586: 1277–1289, 2008. doi: 10.1113/JPHYSIOL.2007.140426.
26. **Martinez-Valdes E, Negro F, Farina D, Falla D.** Divergent response of low- versus high-threshold motor units to experimental muscle pain. *J Physiol* 598: 2093–2108, 2020. doi: 10.1113/JP279225.
27. **Del Santo F, Gelli F, Spidalieri R, Rossi A.** Corticospinal drive during painful voluntary contractions at constant force output. *Brain Res* 1128: 91–98, 2007. doi: 10.1016/J.BRAINRES.2006.09.039.

28. **Schabrun SM, Christensen SW, Mrachacz-Kersting N, Graven-Nielsen T.** Motor Cortex Reorganization and Impaired Function in the Transition to Sustained Muscle Pain. *Cerebral Cortex* 26: 1878–1890, 2016. doi: 10.1093/CERCOR/BHU319.
29. **Farina S, Valeriani M, Rosso T, Aglioti S, Tamburin S, Fiaschi A, Tinazzi M.** Transient inhibition of the human motor cortex by capsaicin-induced pain. A study with transcranial magnetic stimulation. *Neurosci Lett* 314: 97–101, 2001. doi: 10.1016/S0304-3940(01)02297-2.
30. **Le Pera D, Graven-Nielsen T, Valeriani M, Oliviero A, Di Lazzaro V, Tonali PA, Arendt-Nielsen L.** Inhibition of motor system excitability at cortical and spinal level by tonic muscle pain. *Clinical Neurophysiology* 112: 1633–1641, 2001. doi: 10.1016/S1388-2457(01)00631-9.
31. **Summers SJ, Chipchase LS, Hirata R, Graven-Nielsen T, Cavaleri R, Schabrun SM.** Motor adaptation varies between individuals in the transition to sustained pain. *Pain* 160: 2115–2125, 2019. doi: 10.1097/J.PAIN.0000000000001604.
32. **Škarabot J, Ammann C, Balshaw TG, Divjak M, Urh F, Murks N, Foffani G, Holobar A, Holobar A.** Decoding firings of a large population of human motor units from high-density surface electromyogram in response to transcranial magnetic stimulation Key points. *J Physiol* 601: 1719–1744, 2023. doi: 10.1113/JP284043#support-information-section.
33. **Zambolin F, Duro Ocana P, Goulding R, Sanderson A, Venturelli M, Wood G, McPhee J, Parr JVV.** The corticomuscular response to experimental pain via blood flow occlusion when applied to the ipsilateral and contralateral leg during an isometric force task. *Psychophysiology* 61: e14466, 2024. doi: 10.1111/PSYP.14466.
34. **Smith SA, Norbury R, Hunt AJ, Mauger AR.** Intra- and interindividual reliability of muscle pain induced by an intramuscular injection of hypertonic saline injection into the quadriceps. *European Journal of Pain* 27: 1216–1225, 2023. doi: 10.1002/EJP.2151.
35. **Rossi S, Hallett M, Rossini PM, Pascual-Leone A.** Screening questionnaire before TMS: An update. *Clinical Neurophysiology* 122: 1686, 2011. doi: 10.1016/J.CLINPH.2010.12.037.
36. **Faul F, Erdfelder E, Lang AG, Buchner A.** G\*Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods* 39: 175–191, 2007. doi: 10.3758/BF03193146/METRICS.
37. **Pullen RL.** Administering medication by the Z-track method. *Nursing (Brux)* 35: 24, 2005. doi: 10.1097/00152193-200507000-00018.
38. **Strojnik V, Komi P V.** Neuromuscular fatigue after maximal stretch-shortening cycle exercise. *J Appl Physiol* 84: 344–350, 1998. doi: 10.1152/JAPPL.1998.84.1.344/ASSET/IMAGES/LARGE/JAPP0511508.JPEG.
39. **Osborne JO, Tallent J, Girard O, Marshall PW, Kidgell D, Buhmann R.** Neuromuscular electrical stimulation during maximal voluntary contraction: a Delphi survey with expert consensus. *Eur J Appl Physiol* 123: 2203–2212, 2023. doi: 10.1007/S00421-023-05232-1/TABLES/2.
40. **Li G, Wang S, Duan YY.** Towards conductive-gel-free electrodes: Understanding the wet electrode, semi-dry electrode and dry electrode-skin interface impedance using electrochemical impedance spectroscopy fitting. *Sens Actuators B Chem* 277: 250–260, 2018. doi: 10.1016/J.SNB.2018.08.155.
41. **Cook DB, O'Connor PJ, Eubanks SA, Smith JC, Lee M.** Naturally occurring muscle pain during exercise: assessment and experimental evidence. *Med Sci Sports Exerc* 29: 999–1012, 1997. doi: 10.1097/00005768-199708000-00004.

- 771 42. **Melzack R.** The short-form McGill pain questionnaire. *Pain* 30: 191–197, 1987. doi:  
772 10.1016/0304-3959(87)91074-8.
- 773 43. **Lackmy A, Marchand-Pauvert V.** The estimation of short intra-cortical inhibition  
774 depends on the proportion of spinal motoneurons activated by corticospinal inputs.  
775 *Clinical Neurophysiology* 121: 612–621, 2010. doi: 10.1016/J.CLINPH.2009.12.011.
- 776 44. **Oya T, Hoffman BW, Cresswell AG.** Corticospinal-evoked responses in lower limb  
777 muscles during voluntary contractions at varying strengths. *J Appl Physiol* 105: 1527–  
778 1532, 2008. doi:  
779 10.1152/JAPPLPHYSIOL.90586.2008/ASSET/IMAGES/LARGE/ZDG011088227000  
780 5.JPEG.
- 781 45. **Bement MKH, Dicapo J, Rasiarmos R, Hunter SK.** Dose response of isometric  
782 contractions on pain perception in healthy adults. *Med Sci Sports Exerc* 40: 1880–  
783 1889, 2008. doi: 10.1249/MSS.0B013E31817EEECC.
- 784 46. **Graven-Nielsen T, McArdle A, Phoenix J, Arendt-Nielsen L, Jensen TS, Jackson  
785 MJ, Edwards RHT.** In vivo model of muscle pain: Quantification of intramuscular  
786 chemical, electrical, and pressure changes associated with saline-induced muscle pain  
787 in humans. *Pain* 69: 137–143, 1997. doi: 10.1016/S0304-3959(96)03270-8.
- 788 47. **Cabral H V., Devecchi V, Oxendale C, Jenkinson N, Falla D, Gallina A.** Effect of  
789 movement-evoked and tonic experimental pain on muscle force production. *Scand J*  
790 *Med Sci Sports* 34: e14509, 2023. doi: 10.1111/SMS.14509.
- 791 48. **Eon P, Jubeau M, Cattagni T.** Post-activation potentiation after isometric  
792 contractions is strongly related to contraction intensity despite the similar torque–time  
793 integral. *Exp Physiol* 109: 915–925, 2024. doi: 10.1113/EP091700.
- 794 49. **Farina D, Arendt-Nielsen L, Graven-Nielsen T.** Experimental muscle pain decreases  
795 voluntary EMG activity but does not affect the muscle potential evoked by  
796 transcutaneous electrical stimulation. *Clinical Neurophysiology* 116: 1558–1565, 2005.  
797 doi: 10.1016/J.CLINPH.2005.03.009.
- 798 50. **Sousa MV, Goethel M, Becker KM, Diefenthaler F, Fernandes RJ, de Santana  
799 Toro Batista I, Vilas-Boas JP, Ervilha U.** Effect of experimentally induced muscle  
800 pain on neuromuscular control of force production. *Hum Mov Sci* 95: 103219, 2024.  
801 doi: 10.1016/J.HUMOV.2024.103219.
- 802 51. **Aagaard P, Simonsen EB, Andersen JL, Magnusson P, Dyhre-Poulsen P.**  
803 Increased rate of force development and neural drive of human skeletal muscle  
804 following resistance training. *J Appl Physiol* 93: 1318–1326, 2002. doi:  
805 10.1152/JAPPLPHYSIOL.00283.2002/ASSET/IMAGES/LARGE/DG1021846008.JP  
806 EG.
- 807 52. **D’Emanuele S, Tarperi C, Rainoldi A, Schena F, Boccia G.** Neural and contractile  
808 determinants of burst-like explosive isometric contractions of the knee extensors.  
809 *Scand J Med Sci Sports* 33: 127–135, 2023. doi: 10.1111/SMS.14244.
- 810 53. **Rice DA, Mannion J, Lewis GN, McNair PJ, Fort L.** Experimental knee pain  
811 impairs joint torque and rate of force development in isometric and isokinetic muscle  
812 activation. *Eur J Appl Physiol* 119: 2065–2073, 2019. doi: 10.1007/S00421-019-  
813 04195-6.
- 814 54. **Del Vecchio A, Negro F, Holobar A, Casolo A, Folland JP, Felici F, Farina D.** You  
815 are as fast as your motor neurons: speed of recruitment and maximal discharge of  
816 motor neurons determine the maximal rate of force development in humans. *J Physiol*  
817 597: 2445–2456, 2019. doi: 10.1113/JP277396.
- 818 55. **Chowdhury NS, Chang WJ, Millard SK, Skippen P, Bilska K, Seminowicz DA,  
819 Schabrun SM.** The Effect of Acute and Sustained Pain on Corticomotor Excitability:

- A Systematic Review and Meta-Analysis of Group and Individual Level Data. *J Pain* 23: 1680–1696, 2022. doi: 10.1016/J.JPAIN.2022.04.012.
56. **Rice DA, Graven-Nielsen T, Lewis GN, McNair PJ, Dalbeth N.** The effects of experimental knee pain on lower limb corticospinal and motor cortex excitability. *Arthritis Res Ther* 17: 1–8, 2015. doi: 10.1186/S13075-015-0724-0/FIGURES/2.
  57. **Rice DA, Lewis GN, Graven-Nielsen T, Luther R, McNair PJ.** Experimental Hand and Knee Pain Cause Differential Effects on Corticomotor Excitability. *J Pain* 22: 789–796, 2021. doi: 10.1016/J.JPAIN.2021.01.006.
  58. **Goodall S, Howatson G, Thomas K.** Modulation of specific inhibitory networks in fatigued locomotor muscles of healthy males. *Exp Brain Res* 236: 463–473, 2018. doi: 10.1007/S00221-017-5142-X/FIGURES/5.
  59. **Mesquita RNO, Škarabot J, Pearcey GEP.** Low-threshold motor units can be a pain during experimental muscle pain. *Journal of Physiology* 598: 2545–2547, 2020. doi: 10.1113/JP279872.
  60. **Martinez-Valdes E, Negro F, Arvanitidis M, Farina D, Falla D.** Pain-induced changes in motor unit discharge depend on recruitment threshold and contraction speed. *J Appl Physiol* 131: 1260–1271, 2021. doi: 10.1152/JAPPLPHYSIOL.01011.2020/ASSET/IMAGES/LARGE/JAPPLPHYSIOL.01011.2020\_F007.JPEG.
  61. **O'Malley CA, Norbury R, Smith SA, Fullerton CL, Mauger AR.** Elevated muscle pain induced by a hypertonic saline injection reduces power output independent of physiological changes during fixed perceived effort cycling. *J Appl Physiol* 137: 99–110, 2024. doi: 10.1152/JAPPLPHYSIOL.00325.2023/ASSET/IMAGES/LARGE/JAPPLPHYSIOL.00325.2023\_F006.JPEG.
  62. **Mense S.** Algesic agents exciting muscle nociceptors. *Exp Brain Res* 196: 89–100, 2009. doi: 10.1007/S00221-008-1674-4/METRICS.
  63. **Hoheisel U, Unger T, Mense S.** Excitatory and modulatory effects of inflammatory cytokines and neurotrophins on mechanosensitive group IV muscle afferents in the rat. *Pain* 114: 168–176, 2005. doi: 10.1016/J.PAIN.2004.12.020.
  64. **Korotkov A, Ljubisavljevic M, Thunberg J, Kataeva G, Roudas M, Pakhomov S, Radovanovic S, Lyskov E, Medvedev S, Johansson H.** Changes in human regional cerebral blood flow following hypertonic saline induced experimental muscle pain: a positron emission tomography study. *Neurosci Lett* 335: 119–123, 2002. doi: 10.1016/S0304-3940(02)01181-3.
  65. **Zambolin F, Laginestra FG, Favaretto T, Giuriato G, Ottaviani MM, Schena F, Duro-Ocana P, McPhee JS, Venturelli M.** Activation of skeletal muscle mechanoreceptors and nociceptors reduces the exercise performance of the contralateral homologous muscles. *Am J Physiol Regul Integr Comp Physiol* 327: R389–R399, 2024. doi: 10.1152/AJPREGU.00069.2024/ASSET/IMAGES/LARGE/AJPREGU.00069.2024\_F003.JPEG.
  66. **Graven-Nielsen T, Jansson Y, Segerdahl M, Kristensen JD, Mense S, Arendt-Nielsen L, Sollevi A.** Experimental pain by ischaemic contractions compared with pain by intramuscular infusions of adenosine and hypertonic saline. *European Journal of Pain* 7: 93–102, 2003. doi: 10.1016/S1090-3801(02)00069-1.
  67. **Goodall S, Thomas K, Barwood M, Keane K, Gonzalez JT, St Clair Gibson A, Howatson G.** Neuromuscular changes and the rapid adaptation following a bout of damaging eccentric exercise. *Acta Physiologica* 220: 486–500, 2017. doi: 10.1111/APHA.12844.



68. **Brownstein CG, Ansdell P, Škarabot J, Howatson G, Goodall S, Thomas K.** An optimal protocol for measurement of corticospinal excitability, short intracortical inhibition and intracortical facilitation in the rectus femoris. *J Neurol Sci* 394: 45–56, 2018. doi: 10.1016/J.JNS.2018.09.001.
69. **Koo TK, Li MY.** A Guideline of Selecting and Reporting Intraclass Correlation Coefficients for Reliability Research. *J Chiropr Med* 15: 155–163, 2016. doi: 10.1016/J.JCM.2016.02.012.
70. **Weidauer L, Zwart MB, Clapper J, Albert J, Vukovich M, Specker B.** Neuromuscular performance changes throughout the menstrual cycle in physically active females. *J Musculoskelet Neuronal Interact* 20: 314, 2020. doi: 10.1249/01.mss.0000486533.03765.e3.
71. **Dragutinovic B, Moser F, Notbohm HL, Ihalainen JK, Bloch W, Schumann M.** Influence of Menstrual Cycle and Oral Contraceptive Phases on Strength Performance, Neuromuscular Fatigue and Perceived Exertion. .

## Figure Captions

**Figure 1.** Schematic of the procedures during visits two and three with the order of these trials being randomised and counterbalanced. Both visits included a. baseline which involved obtaining  $M_{max}$ , determining MVC for subsequent TMS stimulations at 20% MVC. b. Measurement of all dependent variables pre-injection. c. post-injection to evaluate the effects of pain induced by a hypertonic saline injection compared to the injection matched control.  $M_{max}$  = maximal M-wave amplitude; MVC = maximal voluntary contraction; TMS = transcranial magnetic stimulation; spTMS = single pulse transcranial magnetic stimulation; ppTMS = paired pulse transcranial magnetic stimulation. AMT = active motor threshold.

**Figure 2.** Illustration of the pain VAS recording device used in the present study. VAS marker position (0-100 mm) is digitally recorded in real-time. Participants adjust the slider on a moment-by-moment basis to indicate current pain intensity.

**Figure 3.** The absolute intensity of pain measured using a visual analogue scale (VAS) following an injection of hypertonic saline (HYP) or isotonic saline (ISO) during a maximal voluntary isometric contraction (MVC) and contractions of 20% MVC during stimulations of

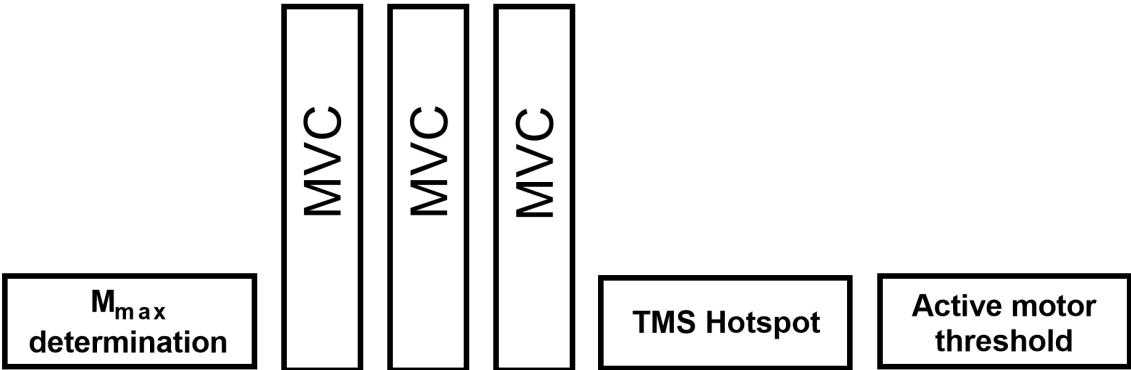
transcranial magnetic stimulation at 120 %, 150% and short interval intracortical inhibition (SICI) of a participant's active motor threshold. Pain intensity values presented were taken the moment before each set was performed. \* denotes significantly different from ISO ( $P < 0.001$ ).

**Figure 4.** Rate of force development calculated as the slope over each 50 ms time period from contraction onset. \* denotes significantly different from HYP (interaction effect).

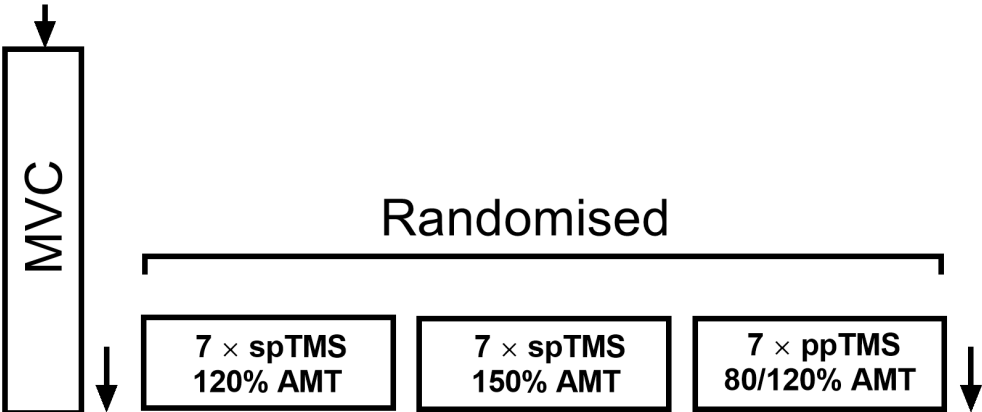
**Figure 5.** Change in neuromuscular function from pre- to post-injection after ISO and HYP. a. Maximal voluntary contraction force. b. Voluntary activation during the MVC. c. Quadriceps potentiated twitch force ( $Q_{tw}$ ). d. M-Wave peak-to-peak amplitude. Data reported as box and whisker plots, with individual changes.

**Figure 6.** TMS responses between ISO and HYP post-injection. a. MEP· $M_{max}$  indicative of corticospinal excitability. b. TMS Silent period duration, indicative of corticospinal inhibition. c. Short interval intracortical inhibition, indicative of cortical inhibition. \* denotes significantly different from ISO in 150% AMT in both VL and RF muscles ( $P < 0.05$ ). Data presented as estimated marginal mean  $\pm$  95% confidence interval. Only significant fixed effects are presented alongside figures.

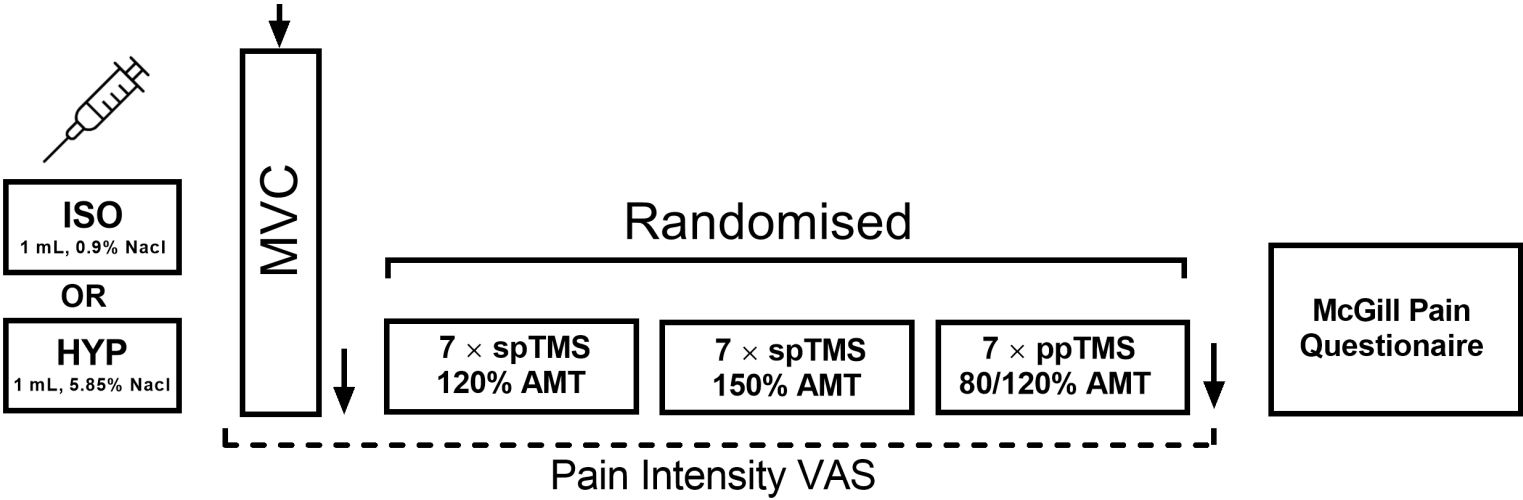
**a. Baseline**

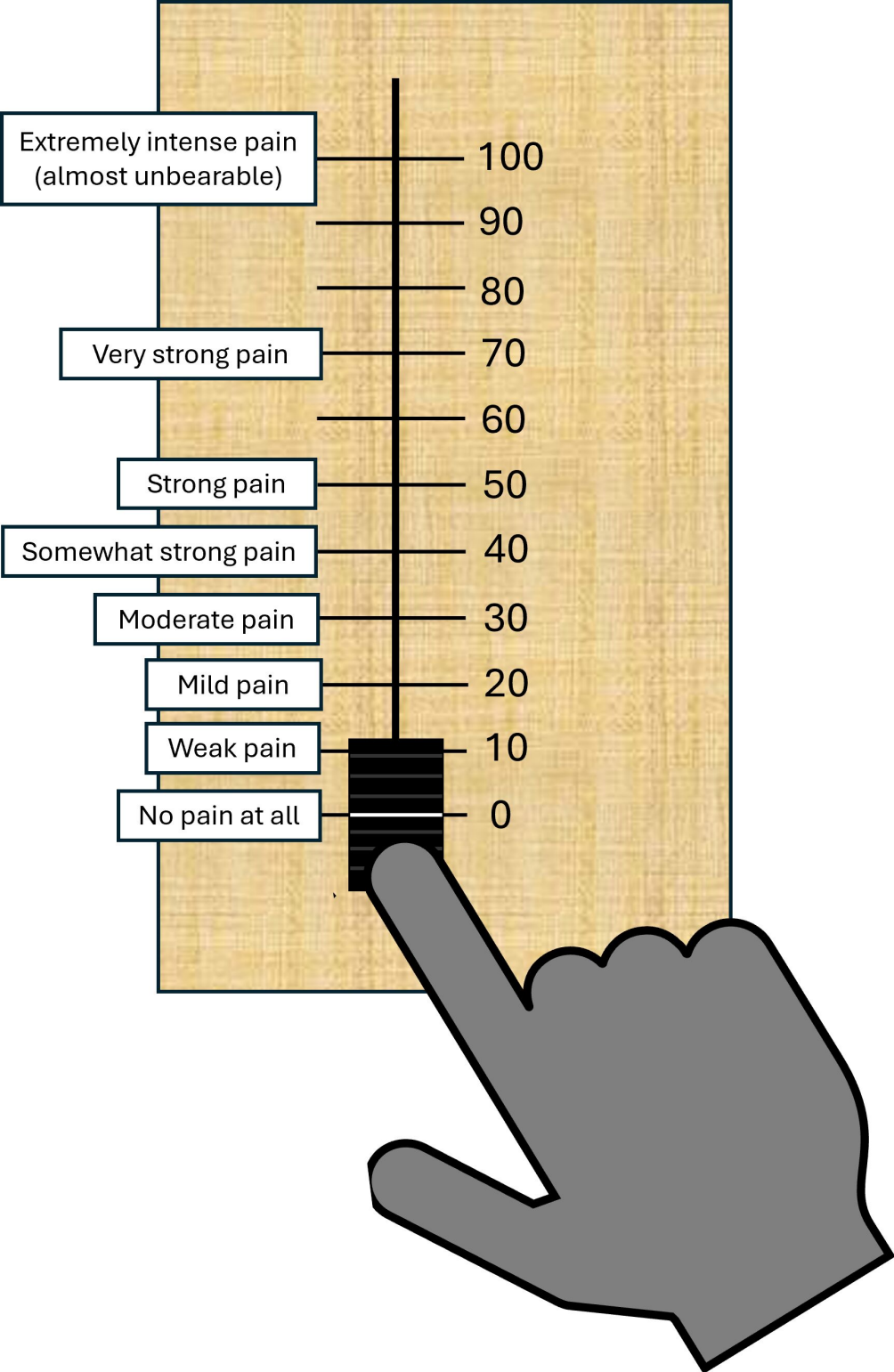


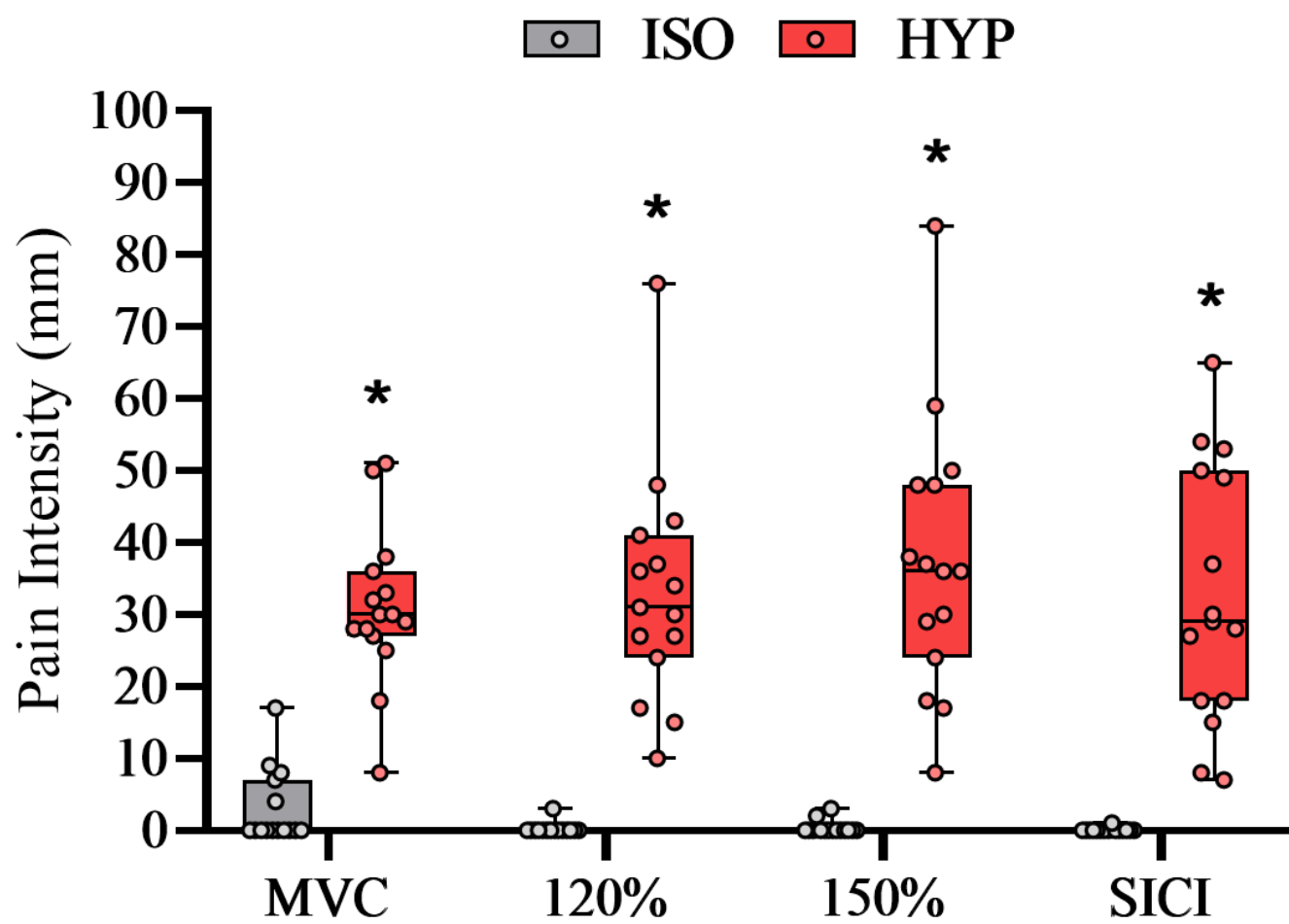
**b. Pre-Injection**

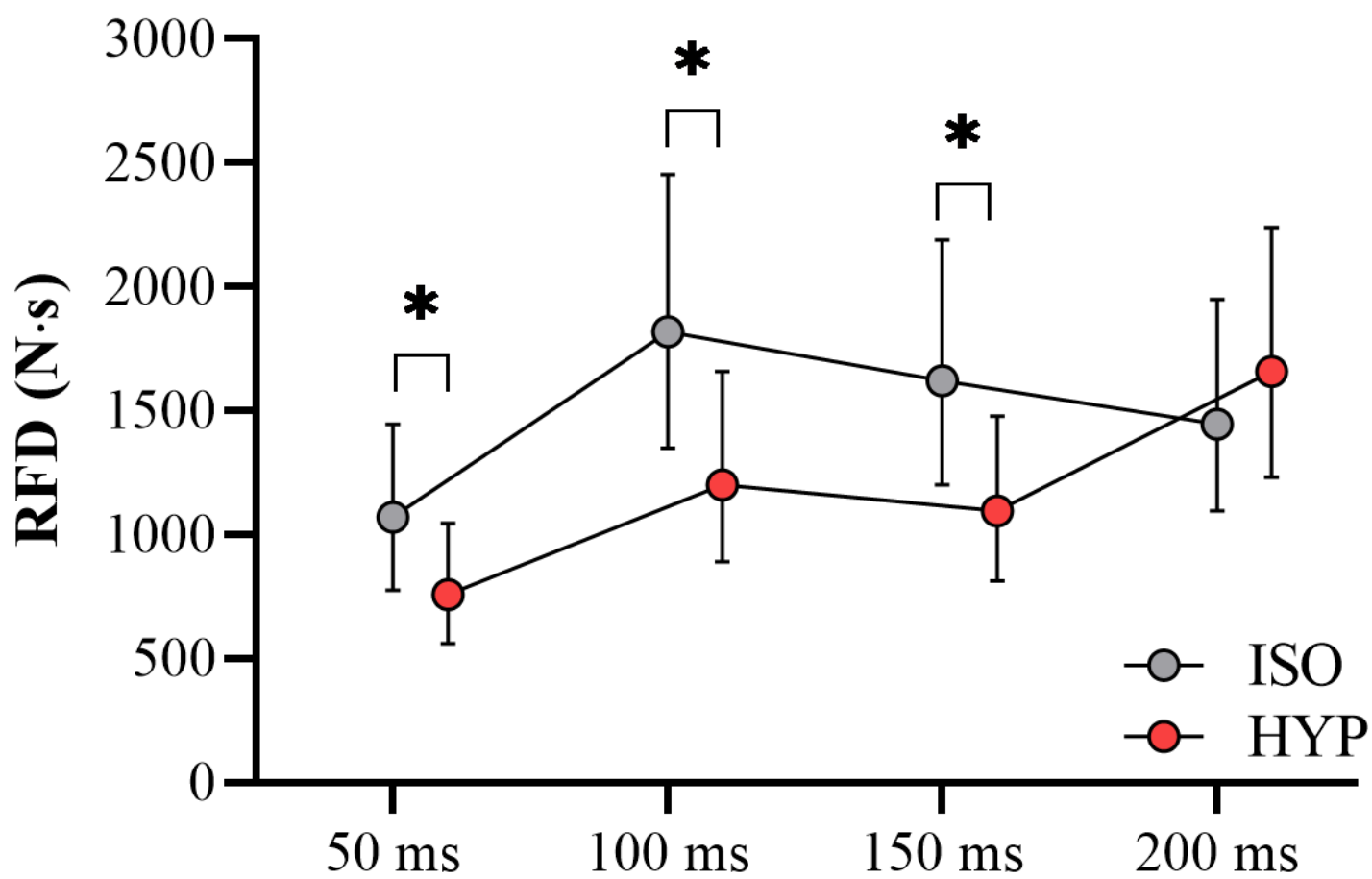


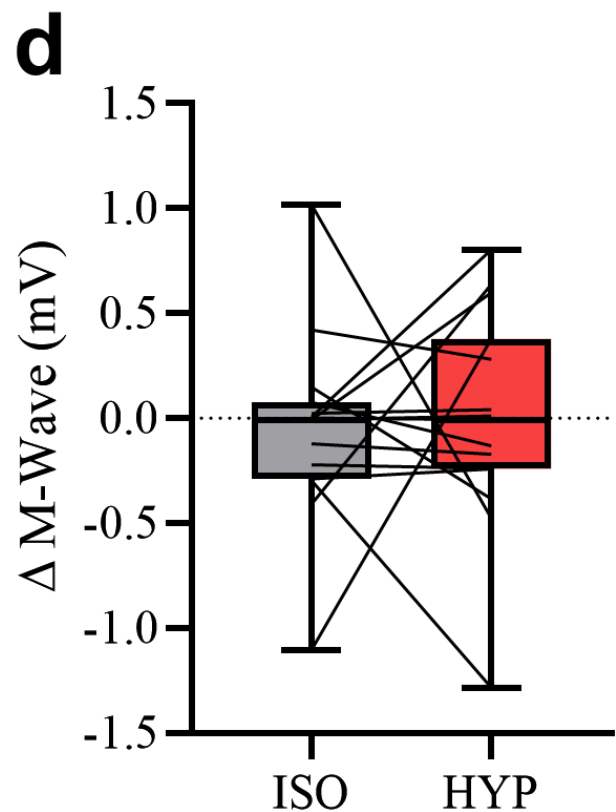
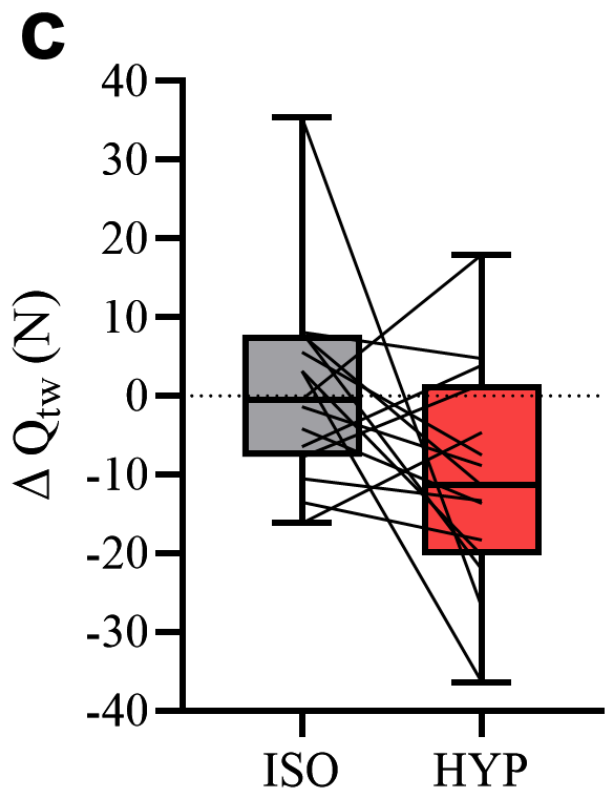
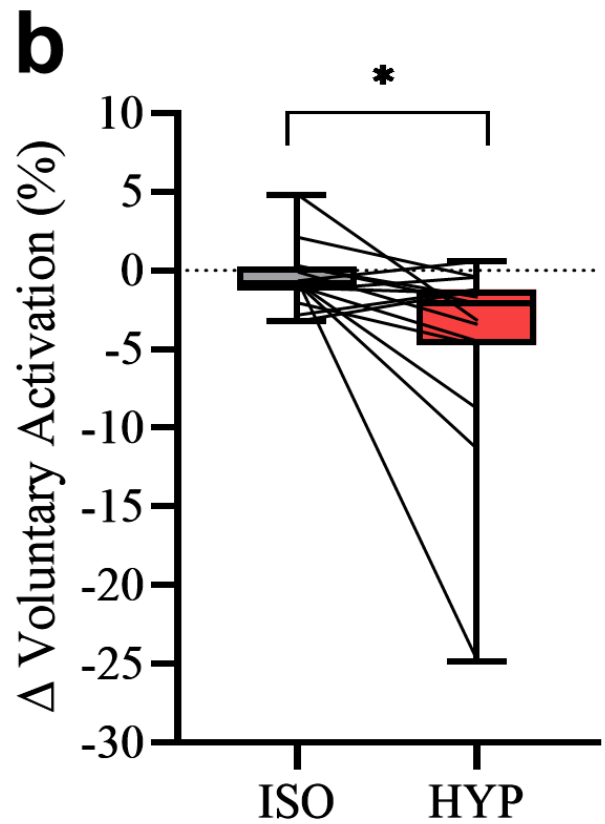
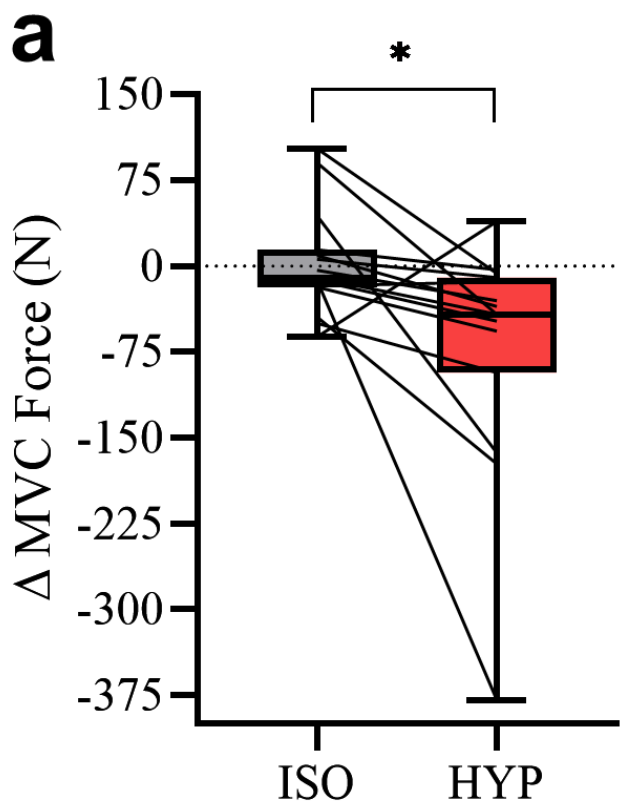
**c. Post-Injection**

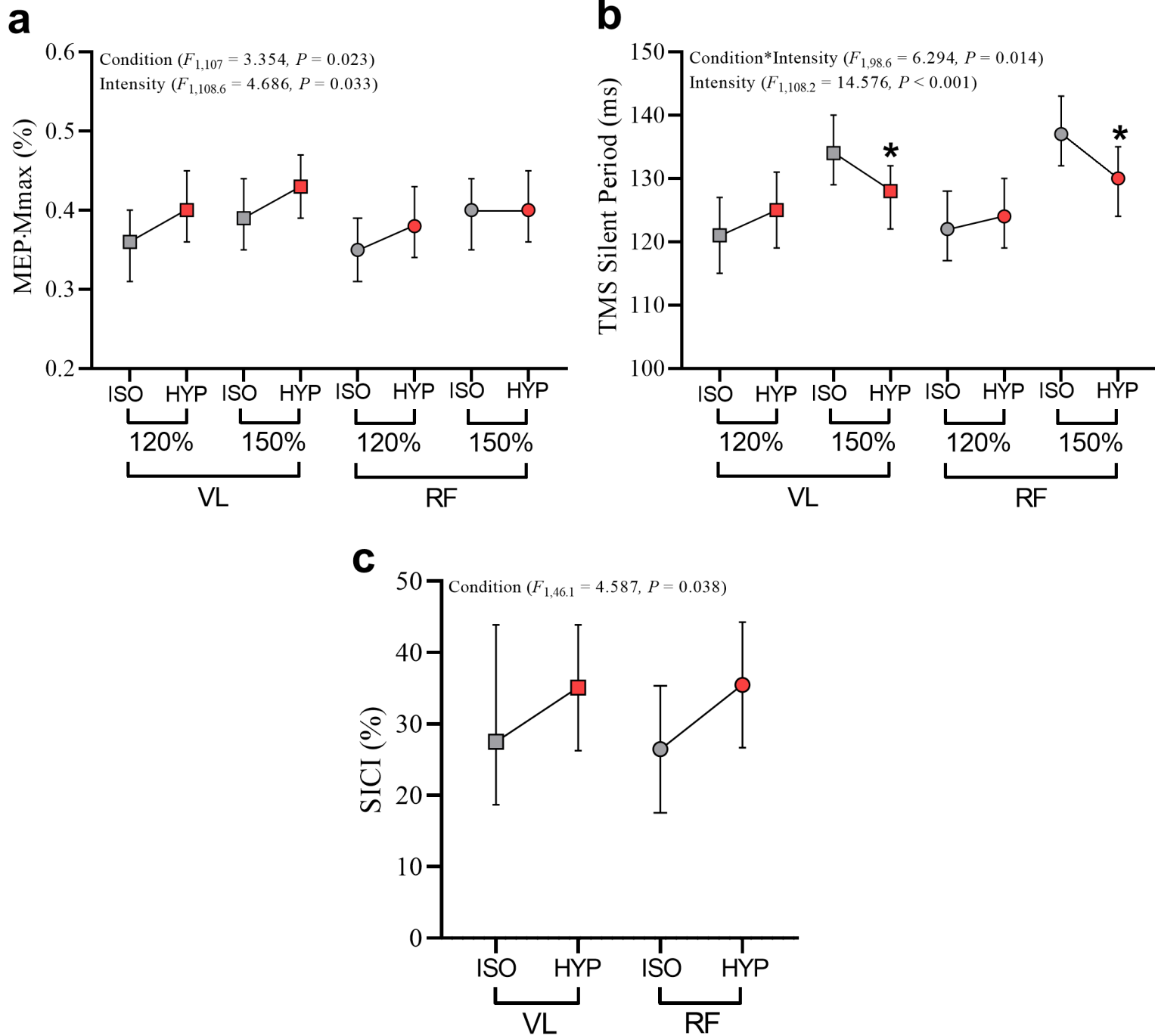










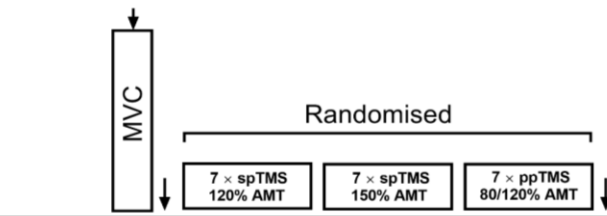




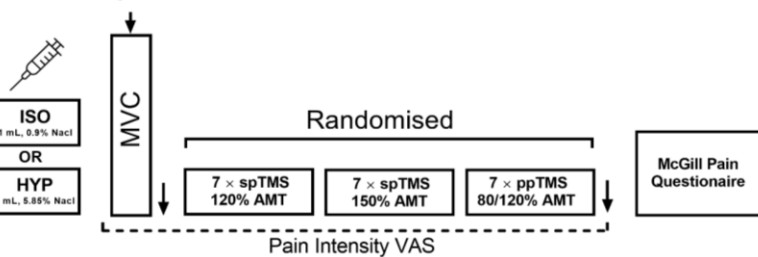
# Experimental muscle pain reduces quadriceps neuromuscular function and alters corticospinal excitability.

## Methods

### Pre-Injection

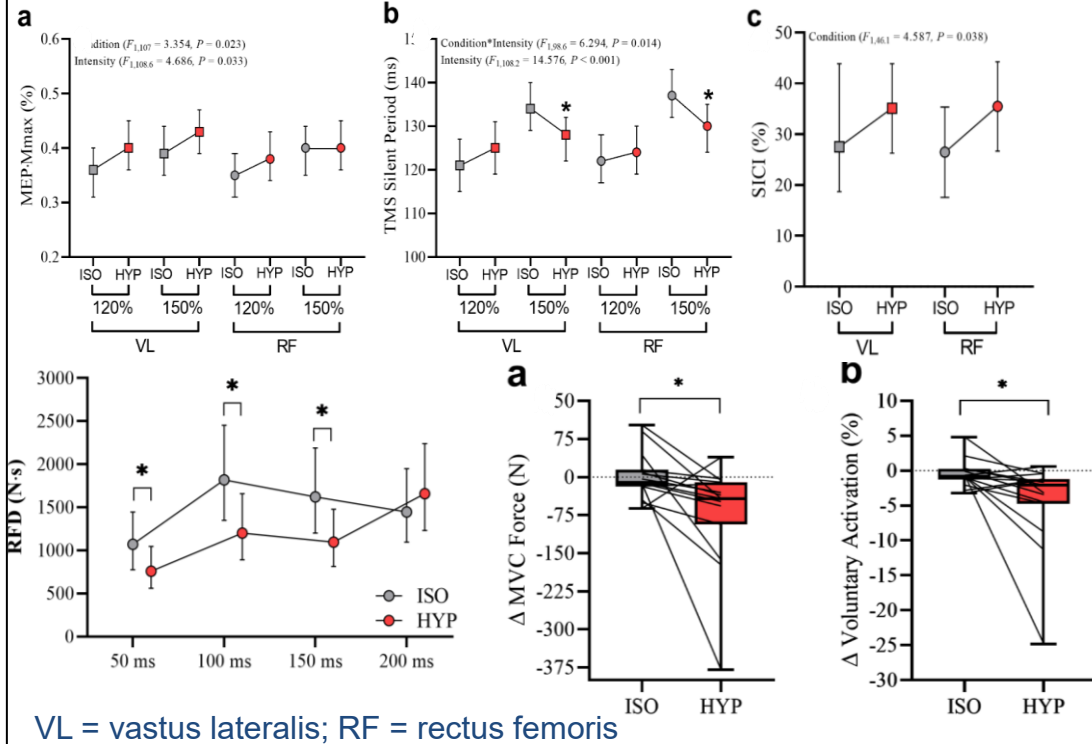


### Post-Injection



Randomised, within-subject design.  
MVC = maximal voluntary contraction; Sp and pp TMS = single/paired pulse transcranial magnetic stimulation.

## Outcomes



## Conclusion

- Increased pain from a hypertonic saline injection (HYP) reduced maximal voluntary force and voluntary activation compared to a non-painful isotonic saline injection (ISO).
- Rate of force development was significantly slower from 50 – 150 ms in the painful, hypertonic saline condition.
- Corticospinal responses show both excitatory and inhibitory responses, with some evidence to suggest stimulation intensity dependent effects.